

Cellular Composition of Tumors

Carcinomas: *Epithelial cell-derived.*

Stroma:

Fibroblasts, Myofibroblasts, Fibrocytes
Inflammatory/Immune Cells

Lymphocytes

T-Cells, Dendritic cells

NK cells

Neutrophils

Monocytes/Macrophages

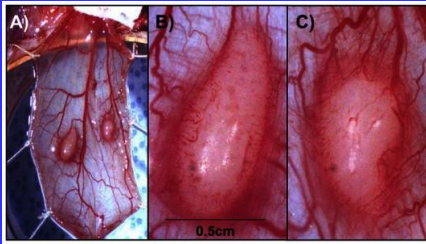
Mast cells

Vascular Cells

Endothelial cells

Endothelial Precursor Cells

Pericytes/Smooth Muscle Cells



Sarcomas: *Mesenchymal cell-derived*

Stroma:

All the above!

Relationship Between Wound Healing and Carcinogenesis

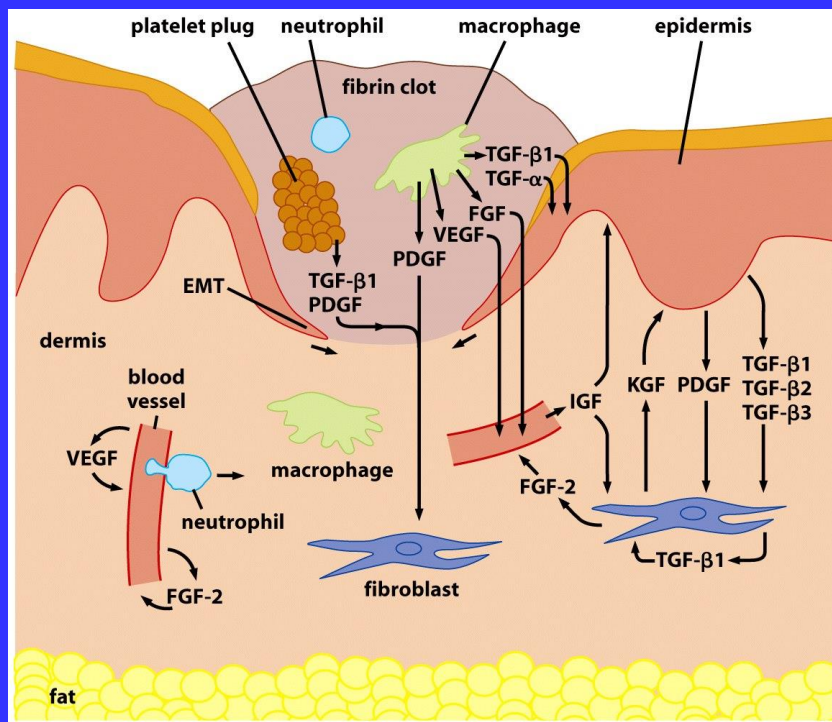


Figure 13.14 *The Biology of Cancer* (© Garland Science 2007)

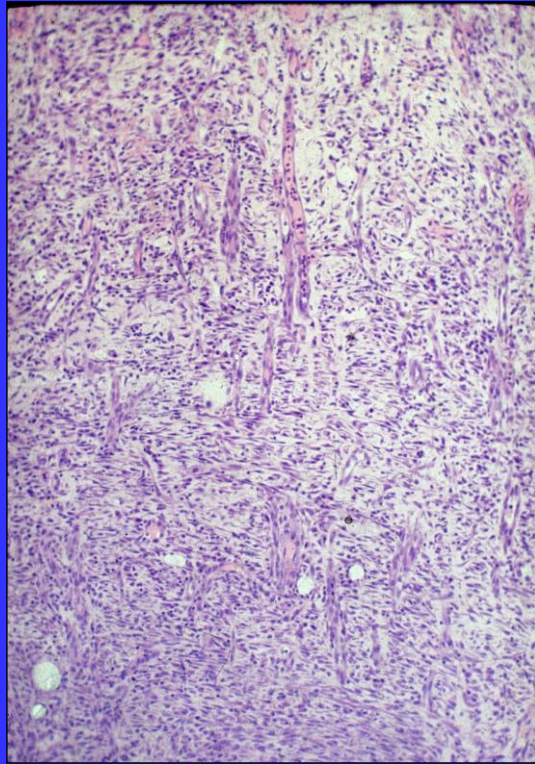


Moses Judah Folkman, 1933-2008
"Father of Angiogenesis"

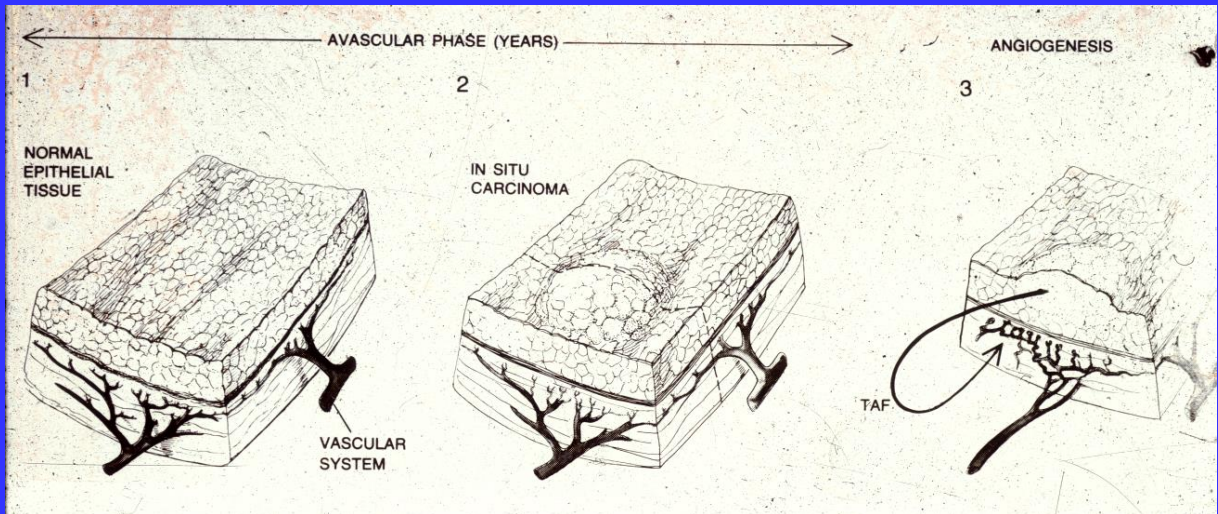
ANGIOGENESIS

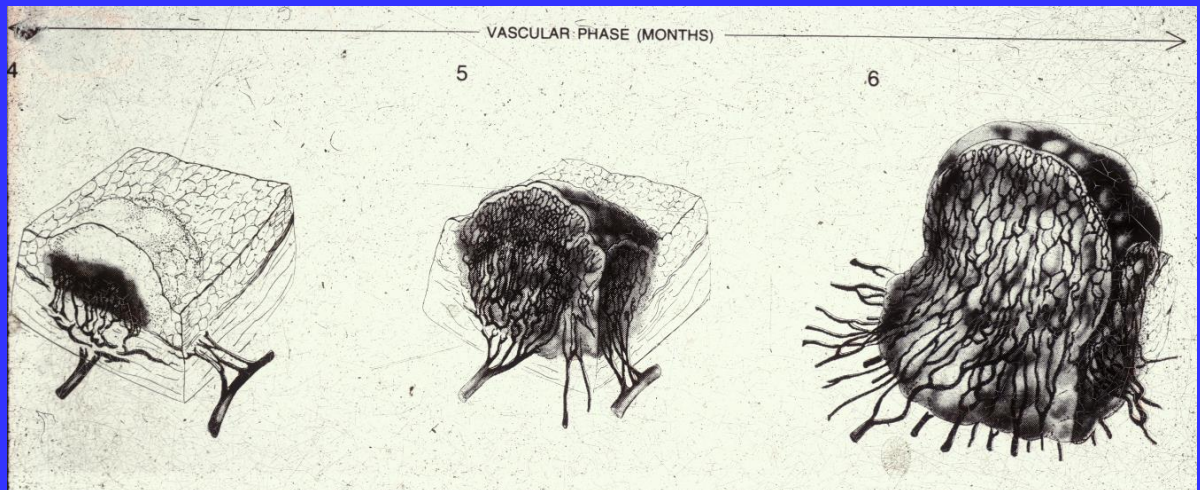
- Prominent during embryogenesis, development and growth
- Virtually absent in adults
- Prominent in ovulation, menstrual cycle and placental formation
- Critical in wound repair and granulation tissue formation
- Prominent in chronic inflammation and fibrosis
- Critical in solid tumor growth and development

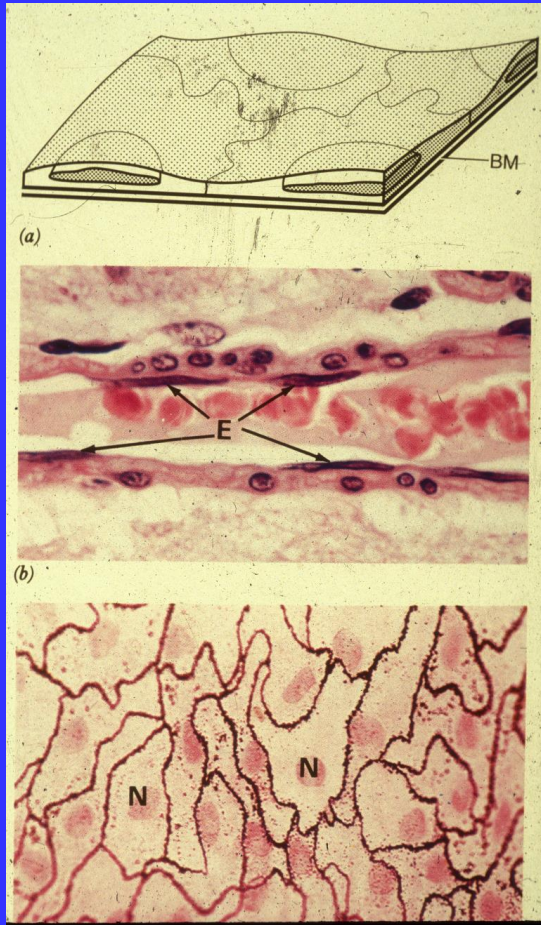
Granulation Tissue



Tumour Angiogenesis







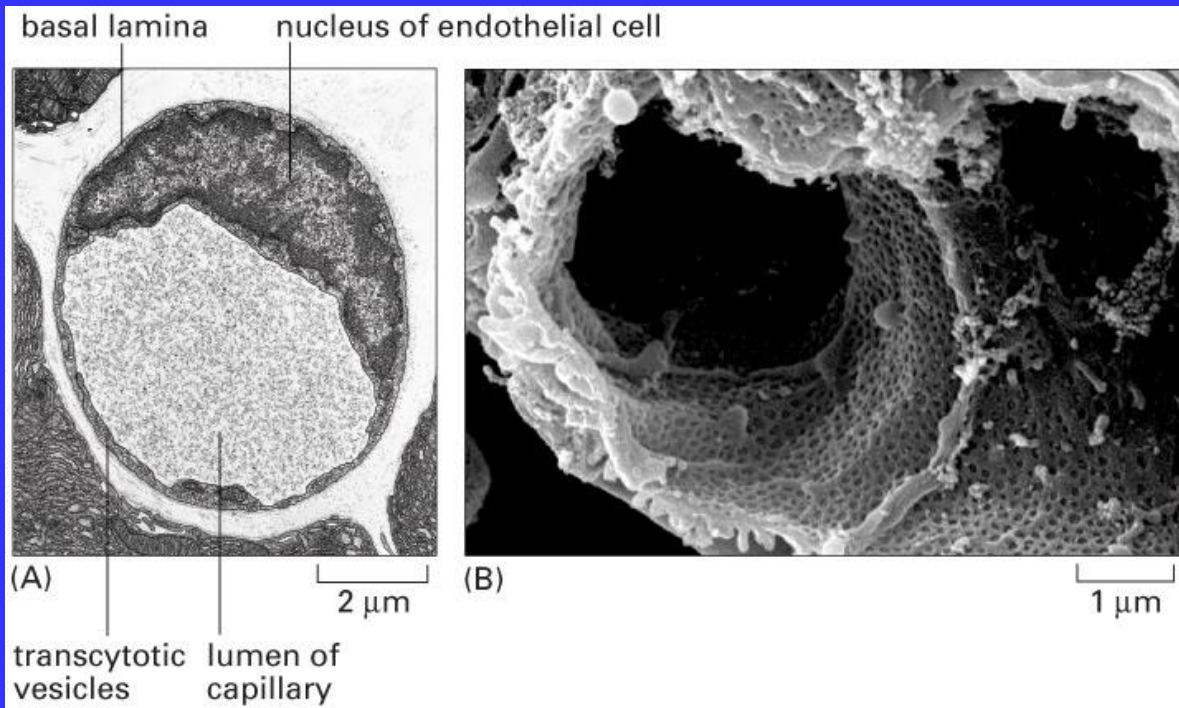


Figure 22-23. Molecular Biology of the Cell, 4th Edition.

ANGIOGENESIS

- Target Cells:
 - Endothelial Cells and Pericytes/Smooth Muscle Cells of Capillaries and Small Venules
- Processes Involved:
 - Disruption of Blood Vessel Continuity
 - Activation/De-Repression of Endothelial Cells
 - Degradation of Basement Membrane
 - Cell Migration
 - Cell Proliferation
 - Lumen Formation
 - Reformation of Basement Membrane
 - Cell Maturation
 - Capillary Loop Formation

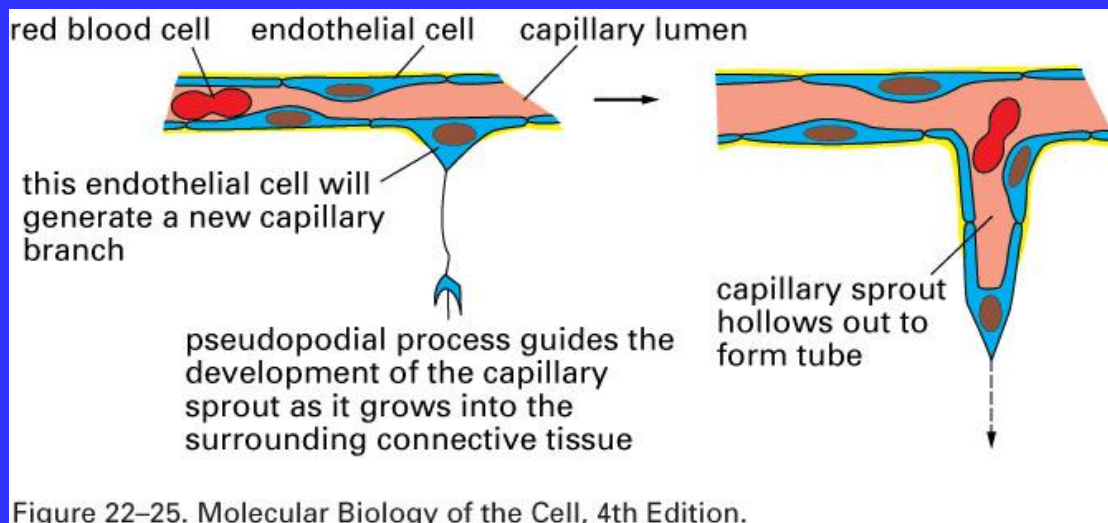
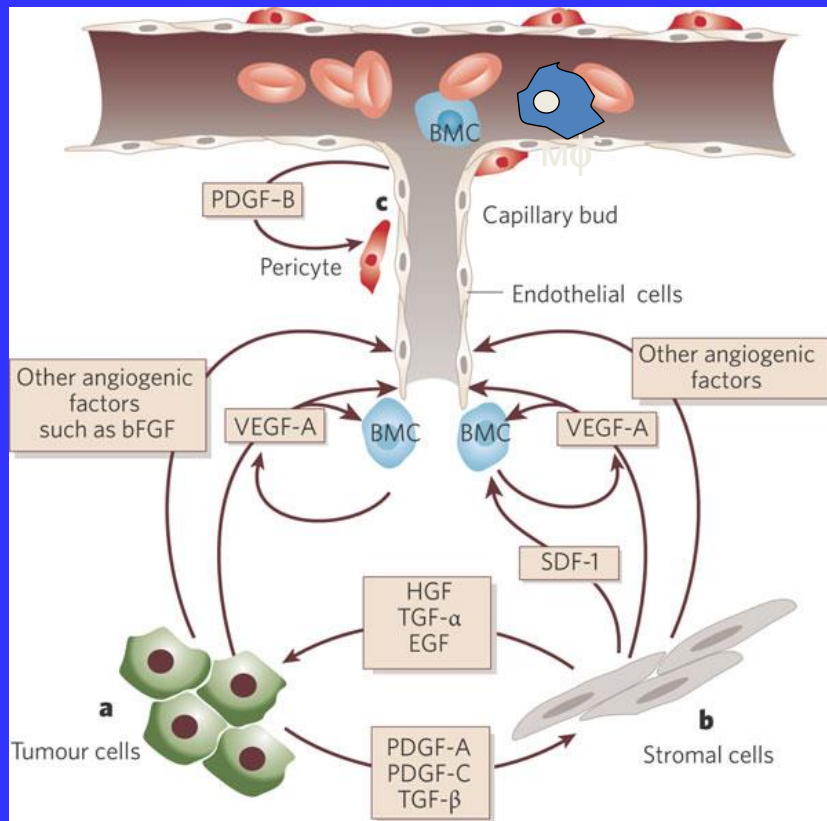


Figure 22-25. Molecular Biology of the Cell, 4th Edition.



ANGIOGENIC FACTORS

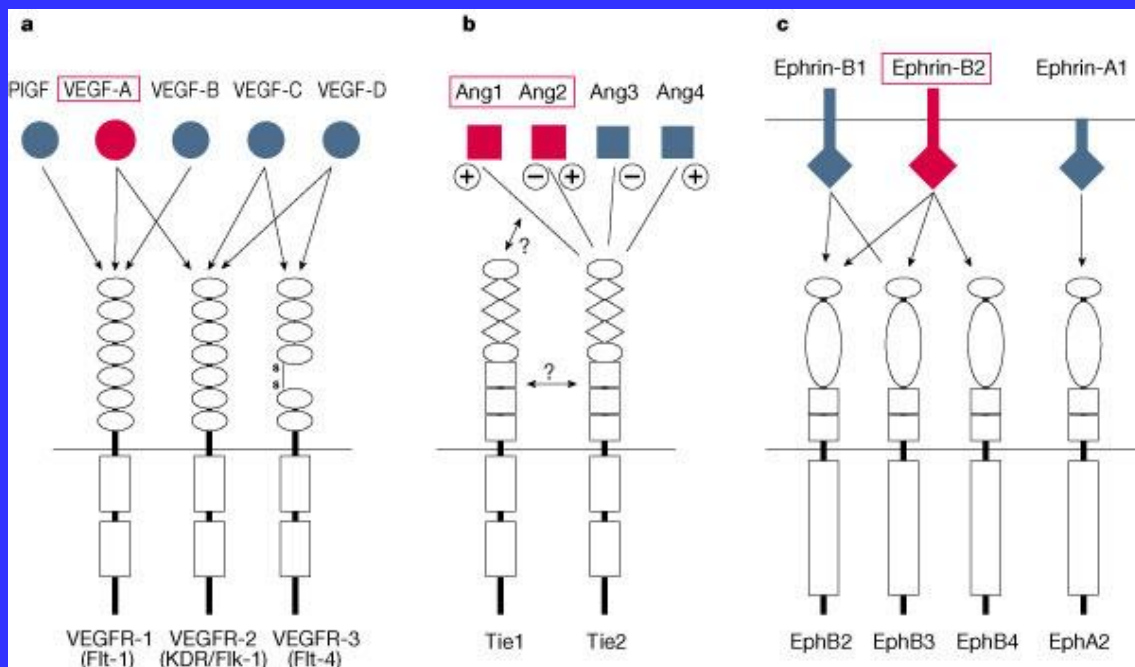
- Fibroblast Growth Factors (FGFs)
 - Two major forms:
 - FGF-1 (aFGF) and FGF-2 (bFGF)
 - M.Wt. 17kDa
 - Bind strongly to HEPARIN
 - No SECRETORY SIGNAL SEQUENCE
 - Found in BASEMENT MEMBRANES
 - Questions:
 - How are FGFs mobilized in Angiogenesis?
 - Are FGFs an autocrine control factor for endothelial cells?

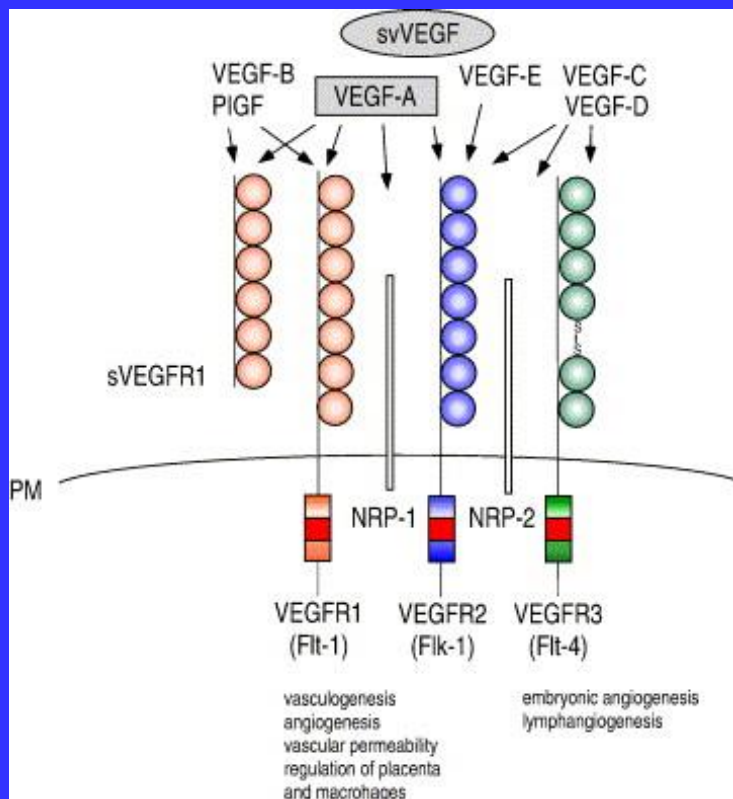
Vascular Endothelial Growth Factor (VEGF) Vascular Permeability Factor

- **Dimeric, heparin binding protein**
- **Potent regulator of vascular permeability**
 - 50,000 X more potent than histamine
- **Secreted by a wide range of tumor cells**
 - carcinomas, sarcomas, glioblastomas, monocytic leukemia cells
- **Secreted by macrophages**
- **VEGF = VPF**
 - One gene, eight alternatively spliced exons
 - In human, four molecular species (121, 165, 189, 206 aa)
 - In mouse, three molecular species (121, 165, 189 aa)
- **Hypoxia is a major regulator of expression via activation of HIF1.**
- **Adenosine signaling is also major regulator of VEGF expression.**

Receptors & Ligands Involved in Regulation of Angiogenesis

<u>LIGAND</u>	<u>RECEPTOR</u>	<u>FUNCTION</u>
VEGF	VEGFR-2 (KDR, Flk-1) VEGFR-1 (Flt-1)	Endothelial Mitogen
Angiopoietin-1 Angiopoietin-2 (acts as antagonist)	TIE-2	Recruitment of accessory cells (SMCs, pericytes)
?	TIE-1	Endothelial cell-cell Interactions
Ephrin-B2 (Arterial)	Eph-B4 (Venous)	Differentiation of arterial vs venous microvasculature
Neuropilins	VEGF-R2	Patterning?



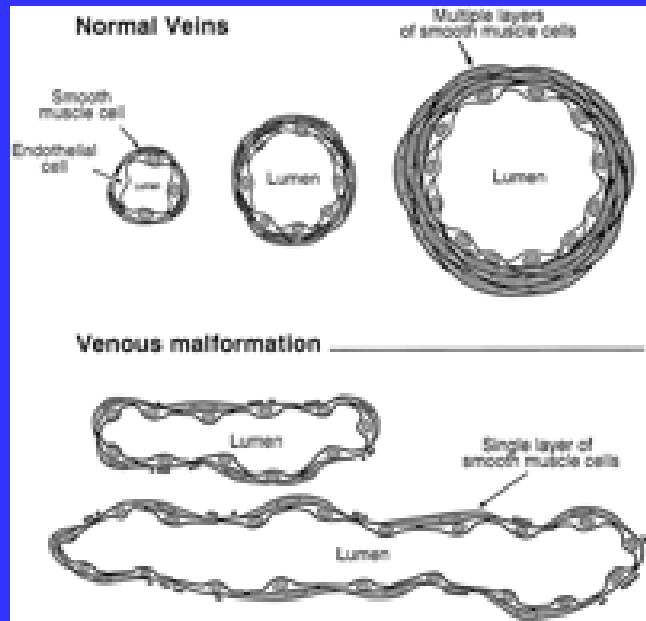


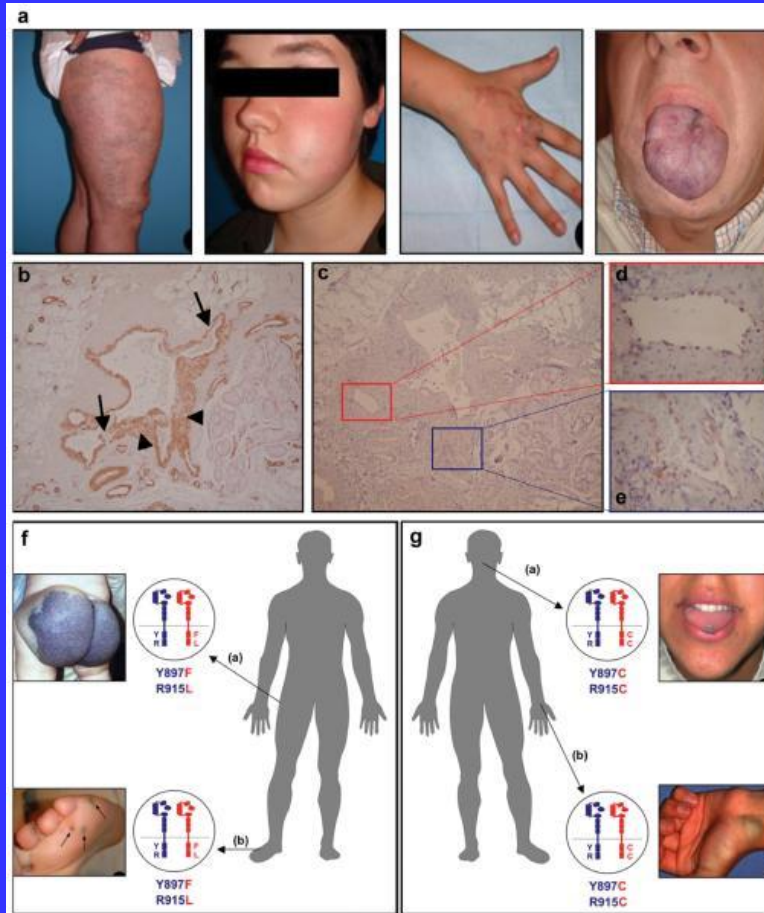
VEGF and VEGF RECEPTORS

- VEGF
 - Essential for both vasculogenesis and angiogenesis
 - Knockouts:
 - Die at 8.5-9.5 days in utero.
 - Delayed differentiation of endothelial cells.
 - Impairment of both angiogenesis and vasculogenesis.
- VEGF-R2 (Flk-1, KDR):
 - Restricted to endothelial cells and their embryonic precursors.
 - Knockouts:
 - Die in utero between 8.5 and 9.5 days.
 - Yolk sac blood islands do not form.
 - No organized blood vessels in embryo yolk sac..
 - Required for hemangioblast to endothelial cell differentiation
- VEGF-R1 (Flt-1):
 - Restricted to endothelial cells and their embryonic precursors.
 - Knockouts:
 - Die in utero at mid-somite stage (Day 9).
 - Essential for organization of embryonic vasculature (EC cell-cell or cell-matrix interaction).
 - Not essential for endothelial cell differentiation.

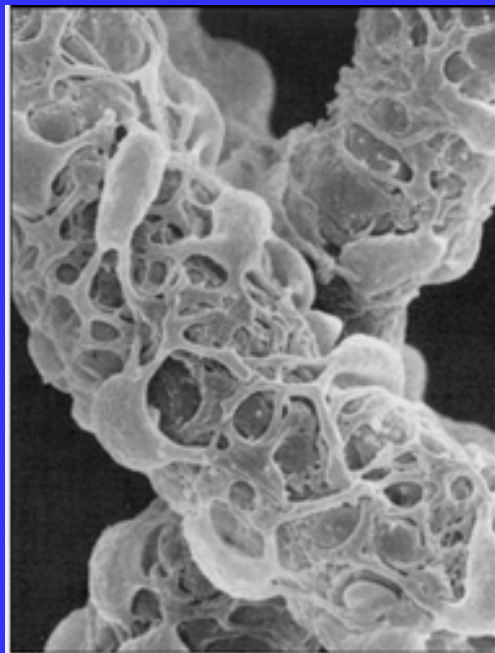
Endothelial Cell-Specific Tyrosine Kinase Receptors

- TIE-1:
 - Knockouts:
 - Form a primitive vasculature
 - Fail to develop structural integrity of vascular endothelial cells.
 - Develop edema and localized hemorrhage.
 - Die in utero at 9-10 days.
 - Ligand unknown
- TIE-2:
 - Knockouts:
 - Fail to recruit smooth muscle cells and pericytes precursors to primitive vasculature.
 - Poorly developed pericardium in heart.
 - Die in utero at 9-10 days.
 - Important in angiogenesis for vascular network formation.
 - Ligand is ANGIOPOIETIN-1 (Ang-1) – EC chemoattractant.
 - Knockouts of Ang-1 have phenotype similar to Tie-2 knockouts.
 - Mutation of TIE-2 in patients (Arg - Tryp):
 - Venous malformations.
 - Develop vein-like structures deficient in non-endothelial cells. Mainly lack smooth muscle cells and pericytes).
 - Angiopietin-2 (Ang-2): Competitive inhibitor of Ang-1.
-





Endothelial Cell - Pericyte Interactions

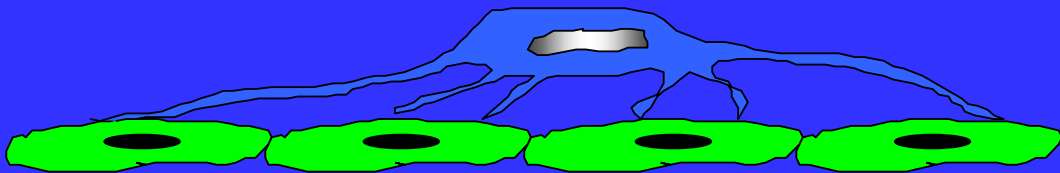


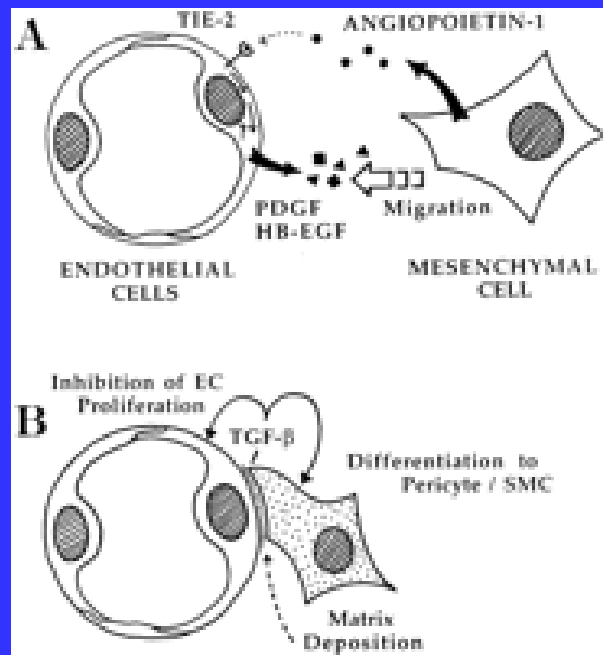
10 μm

Figure 22-24. Molecular Biology of the Cell, 4th Edition.

Endothelial Cell - Pericyte Interactions

- **Recruitment of pericytes involves Tie-2, PDGF and Angiopoietin.**
- Pericytes are associated with the microvasculature, lying outside the endothelium, but within the basement membrane
- Pericytes extend long processes from their cell body, making contact with several endothelial cells
- Interaction is suppressive, resulting in quiescence of endothelial cells
- Activation of latent TGF- β to active TGF- β occurs at sites of pericyte-endothelial cell contact





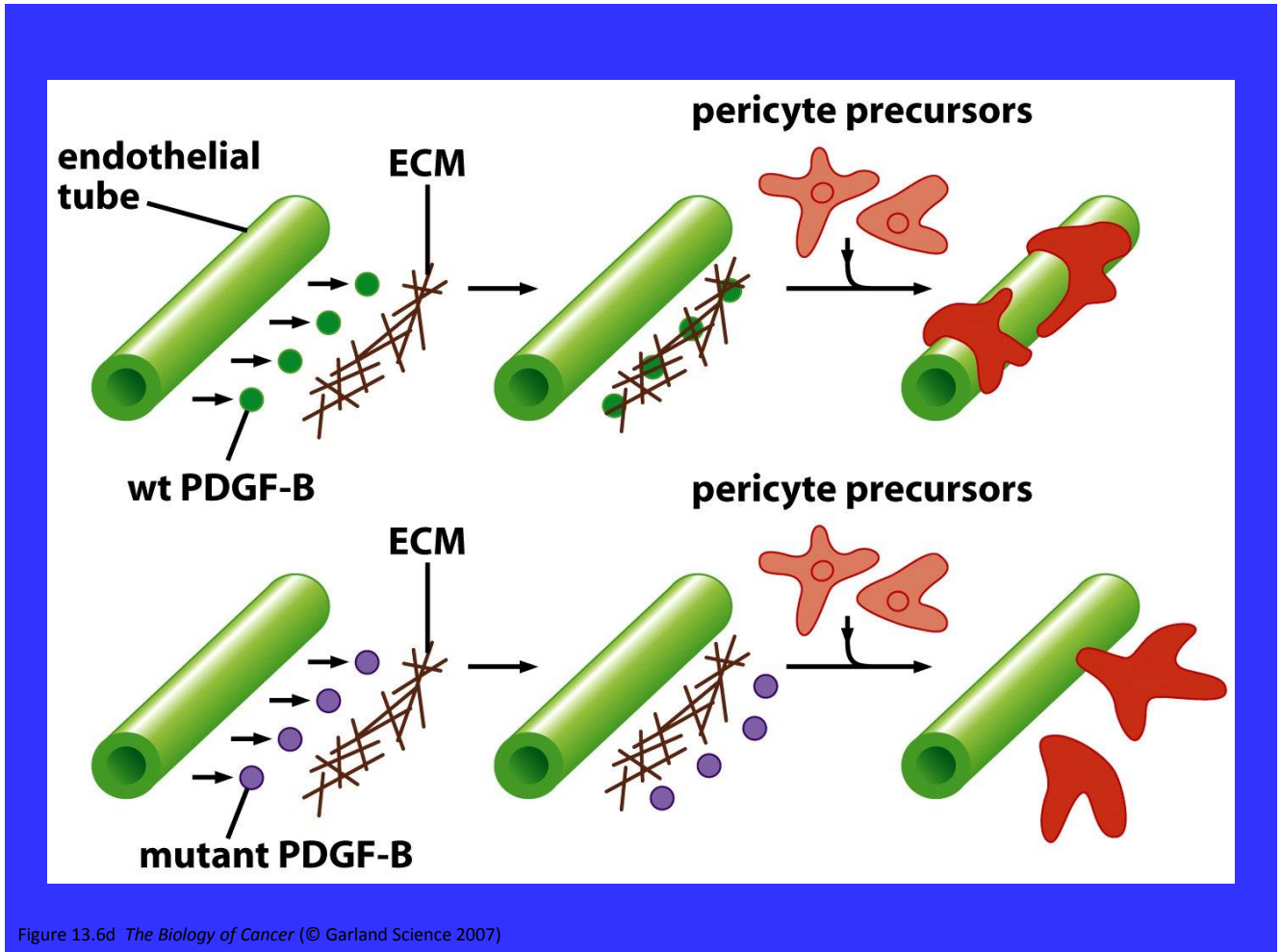


Figure 13.6d *The Biology of Cancer* (© Garland Science 2007)

ARTERIES vs VEINS

Role of Receptor Tyrosine Kinases of the EPHRIN-B / EphB Family

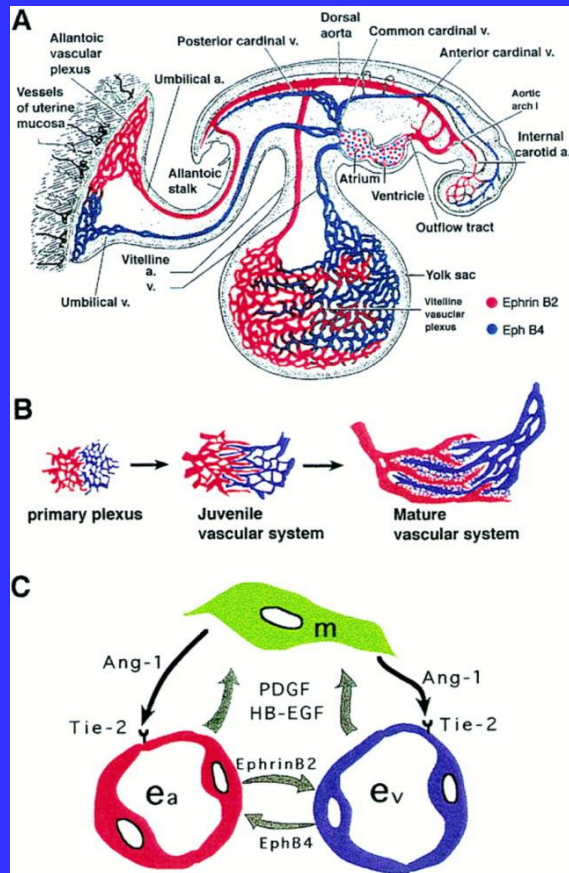
Eph Family: Has at least 14 members

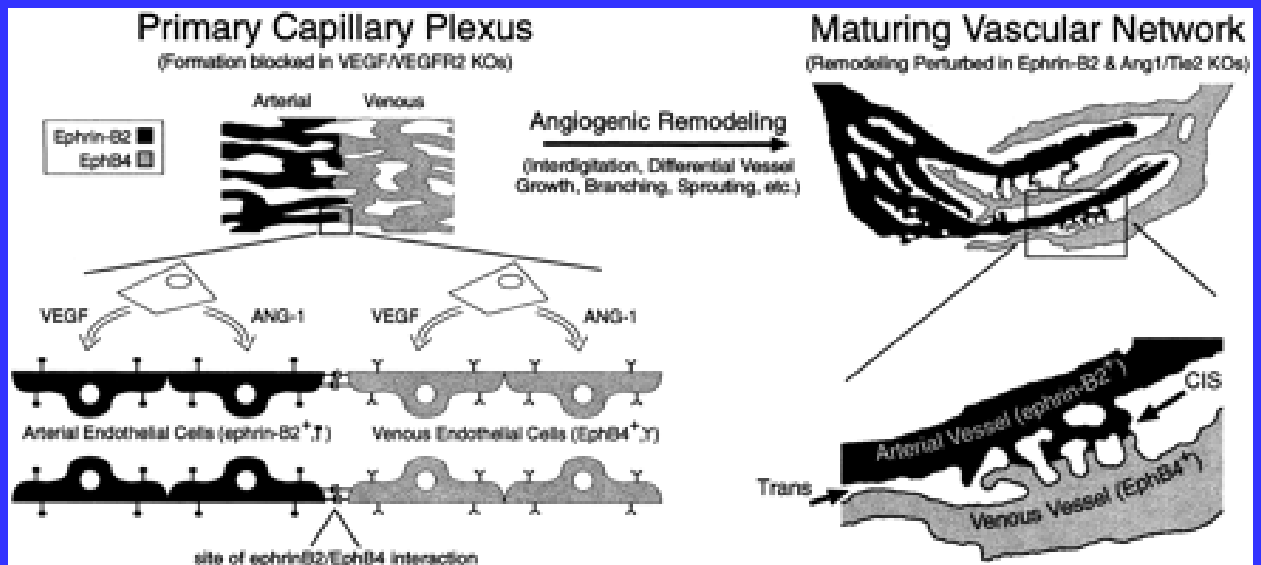
Ephrins: Ligands for Eph family proteins;
At least 8 family members
2 classes: A and B

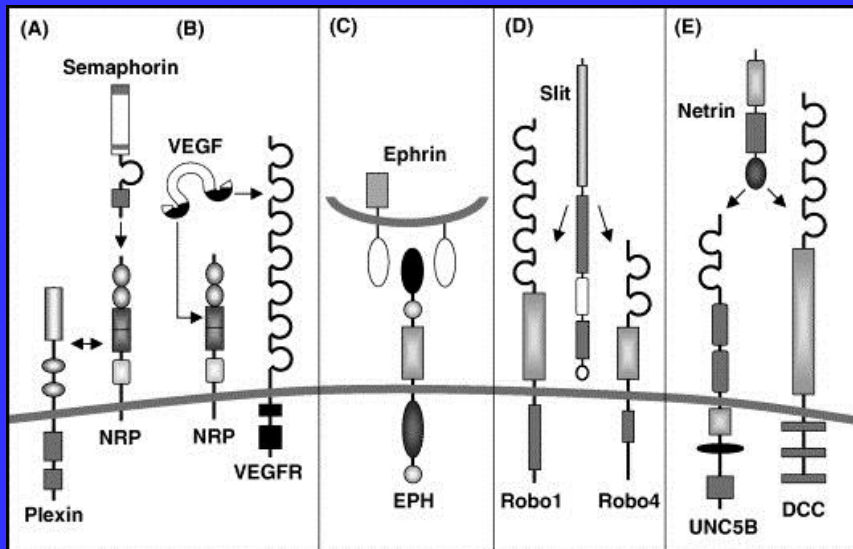
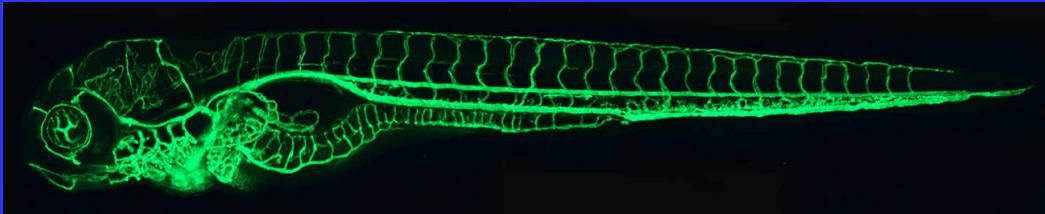
Ephrin-B ligands are trans-membrane proteins, that bind preferentially to receptors of the Eph-B sub-class. Ephrin-A ligands are GPI-linked membrane proteins.

These molecules are NOT SOLUBLE mediators. They are membrane-bound, and activate cognate receptor on partner cells by cell-cell interactions. Signaling is RECIPROCAL, i.e. forward and reverse.

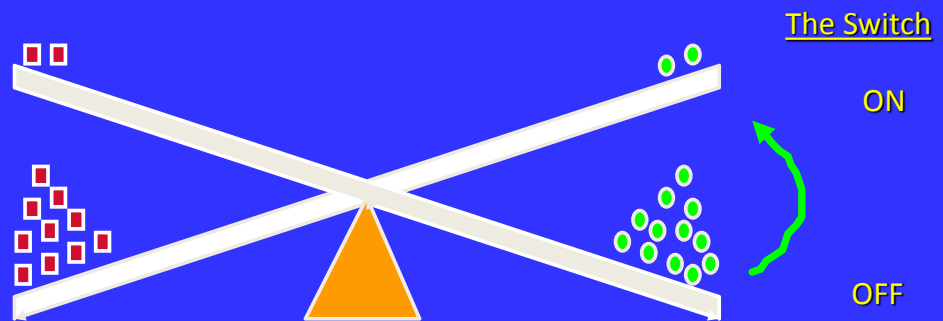
Ephrin-B2 is ARTERIAL. Eph-B4 is Venous







THE BALANCE HYPOTHESIS FOR THE ANGIOGENIC SWITCH



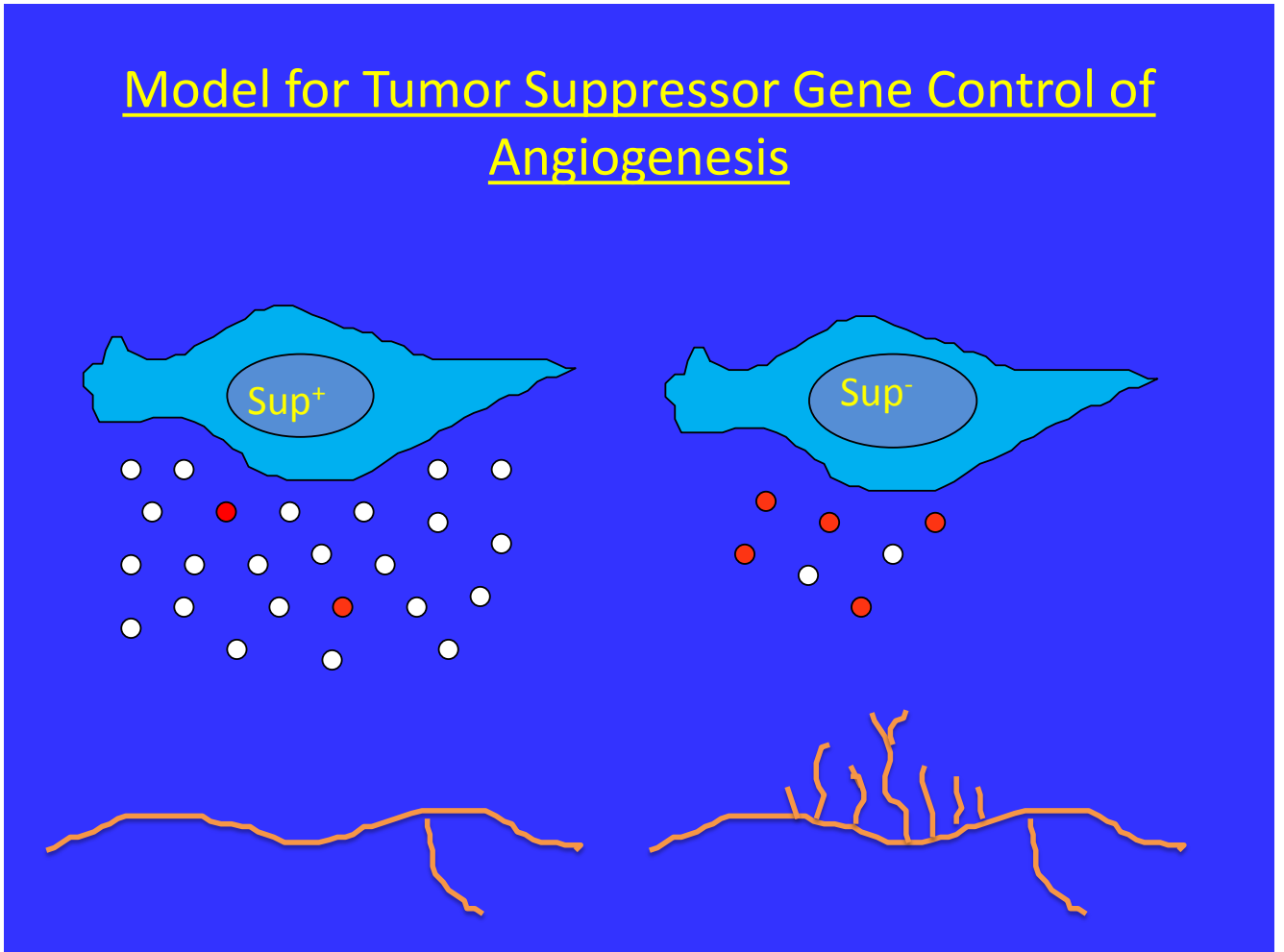
■ ACTIVATORS

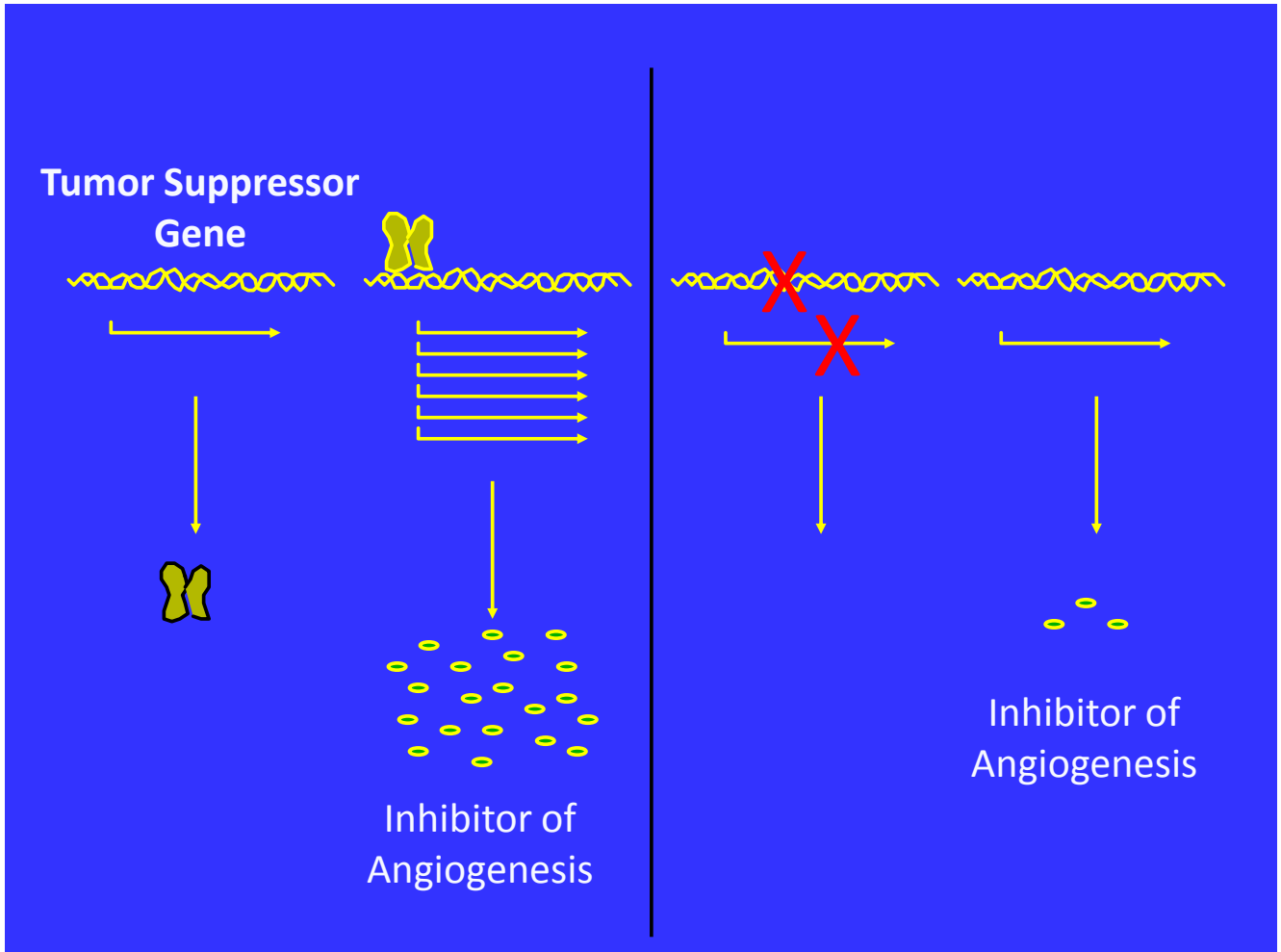
VEGF
 α FGF
bFGF
IL-8

○ INHIBITORS

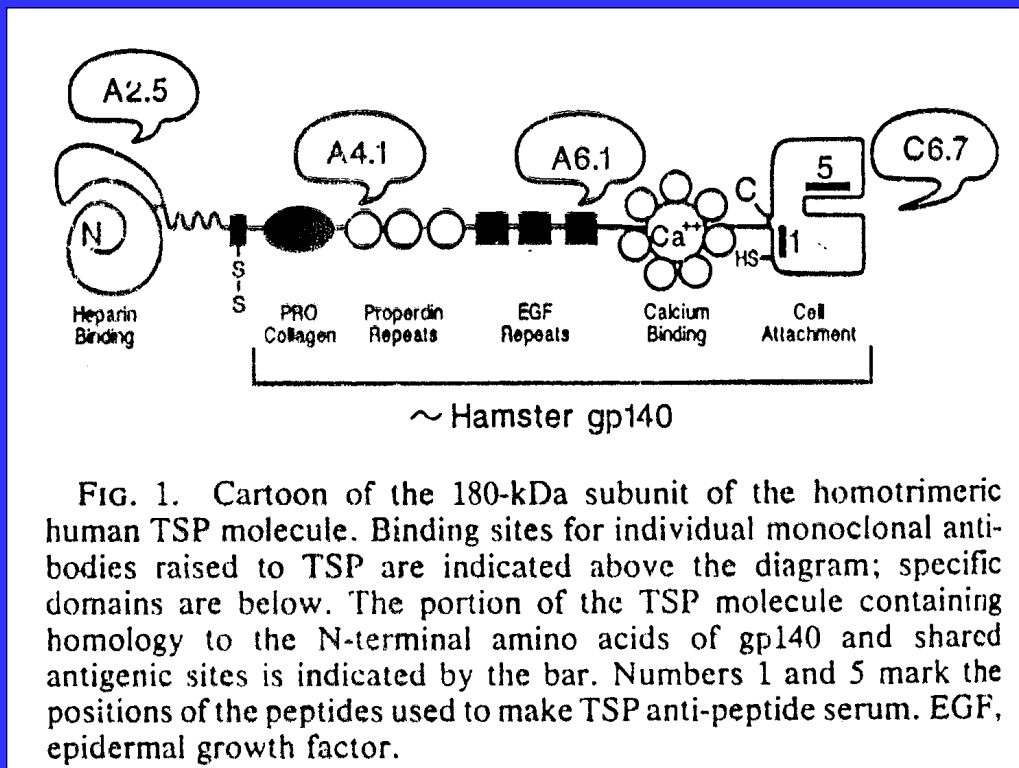
Thrombospondin
TGF β
Angiostatin
Endostatin
TIMP
IL-12
 γ P10

Model for Tumor Suppressor Gene Control of Angiogenesis





Thrombospondin



Li-Fraumeni Fibroblasts and Regulation of Angiogenesis by p53

- LF patients show a greatly increased susceptibility to tumor development.
- Patients have 1 defective p53 allele and 1 wild-type allele.
- LF fibroblasts in culture lose remaining wt p53 allele over several generations.

EARLY PASAGE		LATE PASSAGE
Intact	p53	Lost
Low	Angiogenic Activity	High
High	TSP-1	Low
Low	VEGF	High

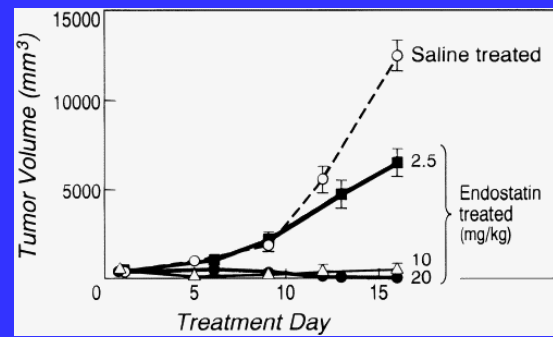
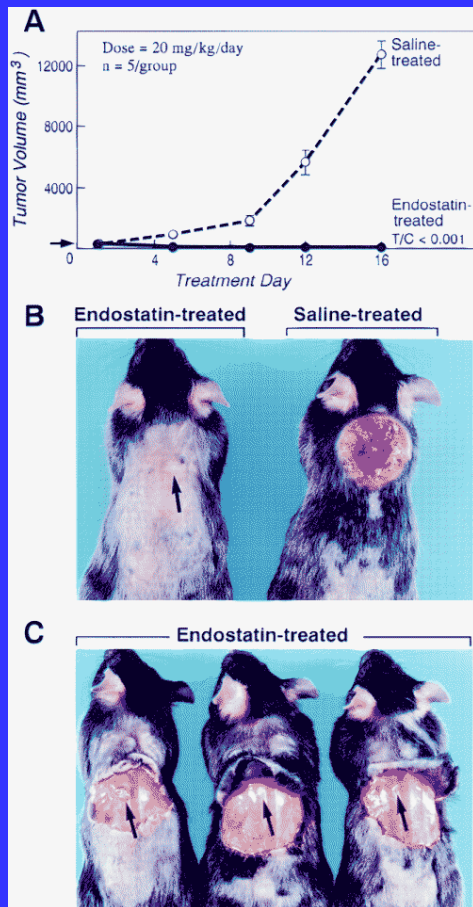
Transfected p53 T⁰-sensitive mutant: WT at 32°C, Mutant at 38°C.

38°C.	TSP-1	Low
32°C.	TSP-1	High

Angiogenesis Inhibitors:

- *ANGIOSTATIN:*
 - Fragment of Plasminogen.
 - Produced by Primary Tumor Mass
 - Present in the Circulation
 - Suppresses Growth of Metastases
 - Removal of Primary Tumor de-suppresses Growth of metastases

- *ENDOSTATIN:*
 - Fragment of Type XVIII collagen



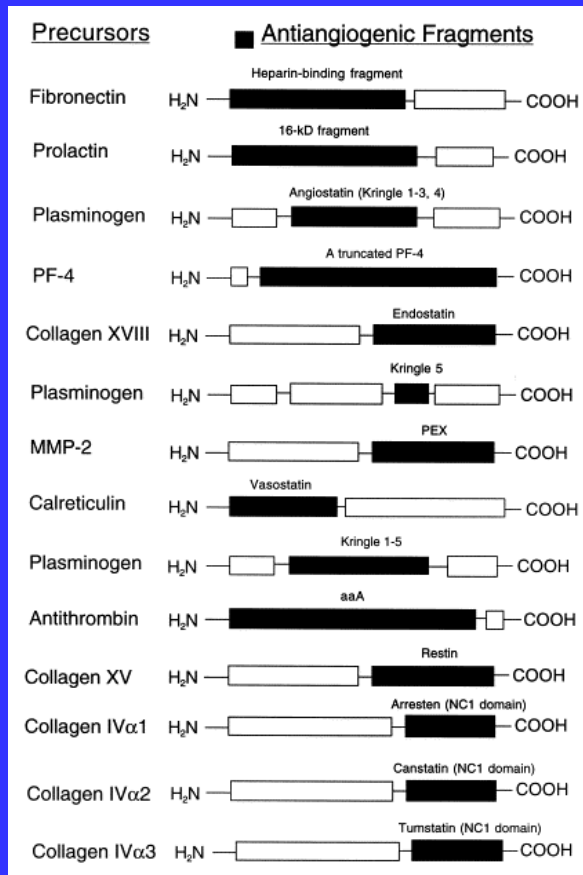


Table 13.4 Angiogenesis inhibitors and their development and use in clinical trials

Name	Status	Responses
A. Endogenous inhibitors of angiogenesis		
Endostatin	in clinical trial	scattered responses
Interferons- α and - β	effective in treating hemangioblastomas	Kaposi's sarcomas; limited efficacy against most other types of tumors
B. Agents that block VEGF and VEGF-R signaling		
Avastin anti-VEGF MoAb	in clinical trial	delayed progression 1–3 months in lung, 3–4 months in colon
SU5416 inhibitor of VEGF-R2 (Flk-1)	trial abandoned	severe vascular toxicities
ZD6474 inhibitor of VEGF-R2	under clinical test	
CP547, 632 inhibitor of VEGF-R2	in trial	
C. Miscellaneous other drugs		
Thalidomide	in trial	inhibits bFGF- and VEGF-dependent angiogenesis
Squalamine sterol from shark liver	in trial	strong anti-angiogenic activity
Celecoxib anti-inflammatory drug	in trial	multiple anti-neoplastic effects
ZD6126	in trial	antagonist of tubulin in endothelial cell cytoskeleton
Fumagillin and TNP-470	in trial; slowed tumor growth	antagonist of methionine aminopeptidase in endothelial cells
D. Inhibitors of ECM breakdown—MMP inhibitors		
Marimastat	in clinical trial	no delay of tumor progression
Prinomastat	in clinical trial	no slowing of tumor progression
BMS275291	in clinical trial	
BAY12-9566	in clinical trial	
Neovastat (shark cartilage MMPI)	in clinical trial	

Table 13.4 *The Biology of Cancer* (© Garland Science 2007)

OXYGEN AND ANGIOGENESIS

- Oxygen Tension & Erythropoiesis
 - Low ppO_2 stimulates **erythropoiesis**
 - Low ppO_2 stimulates **erythropoietin** production in kidney
- Oxygen Tension & Angiogenesis:
 - High ppO_2 inhibits **angiogenesis**
 - Low ppO_2 stimulates **angiogenesis**

Relationship to ALTITUDE PHYSIOLOGY:

- Increased capillary density in muscle at high altitudes. Effect only up to certain elevation.
- FAILURE of WOUND REPAIR above 15,000ft.
 - » (Himalayas Expeditions)



Hypoxia Inducible Factor (HIF1)
Hypoxia Response Element (HRE)

Hypoxia Inducible Factor (HIF)

Dimer of **HIF1- α** and **HIF1- β** (ARNT)

Unstable under Normoxia
Stabilized under hypoxia

Constitutively Expressed
Stable

Mammalian bHLH-PAS proteins

Class I

AHR
CLOCK
HIF-1 α
HIF-2 α (EPAS1/HLF/HRF/MOP2)
HIF-3 α
NPAS1 (MOP5)
NPAS2 (MOP4)
SIM1
SIM2

Class II

ARNT (HIF-1 β)
ARNT2
ARNT3 (BMAL1/MOP3)

Hypoxia Inducible Genes

- Erythropoietin (Epo)
- VEGFs
- Glycolytic Enzymes
 - Lactate Dehydrogenase (LDH)
 - Pyruvate Kinase M (PKMP)
 - Enolase 1
 - Phosphoglycerate Kinase 1 (PGK1)
 - Aldolase A (ALDA)
 - Phosphofructokinase (PFKL)
 - Glucose Transporter 1 (GLUT-1)
- Inducible Nitric Oxide Synthase (iNOS)
- Heme Oxygenase
- Ferritin Receptor and Ferritin
- Tyrosine Hydroxylase

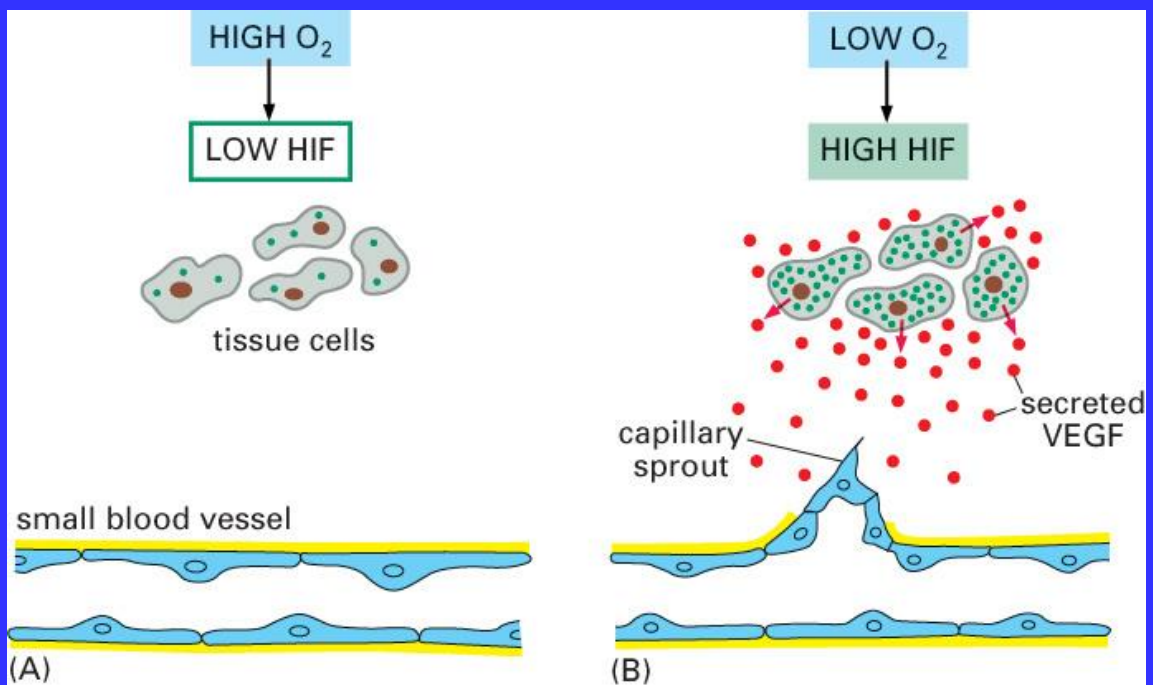
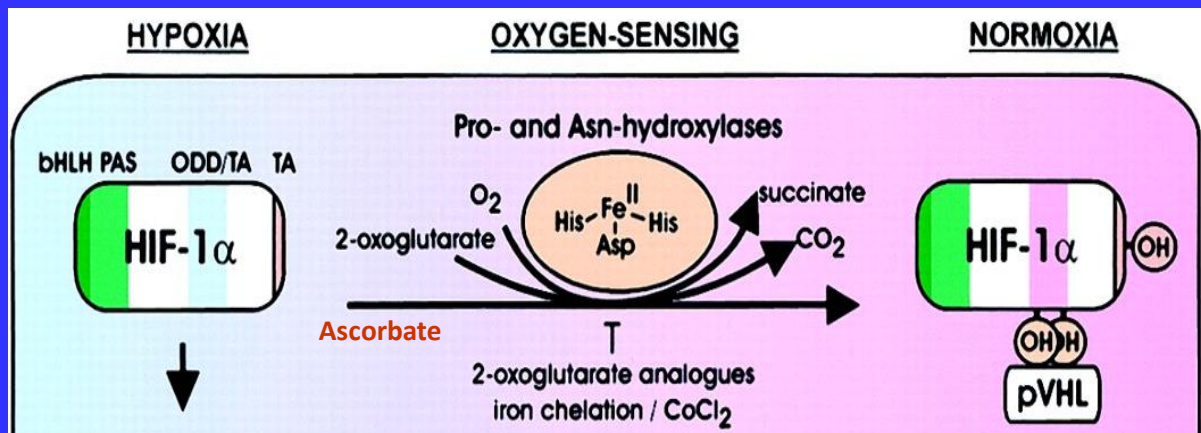
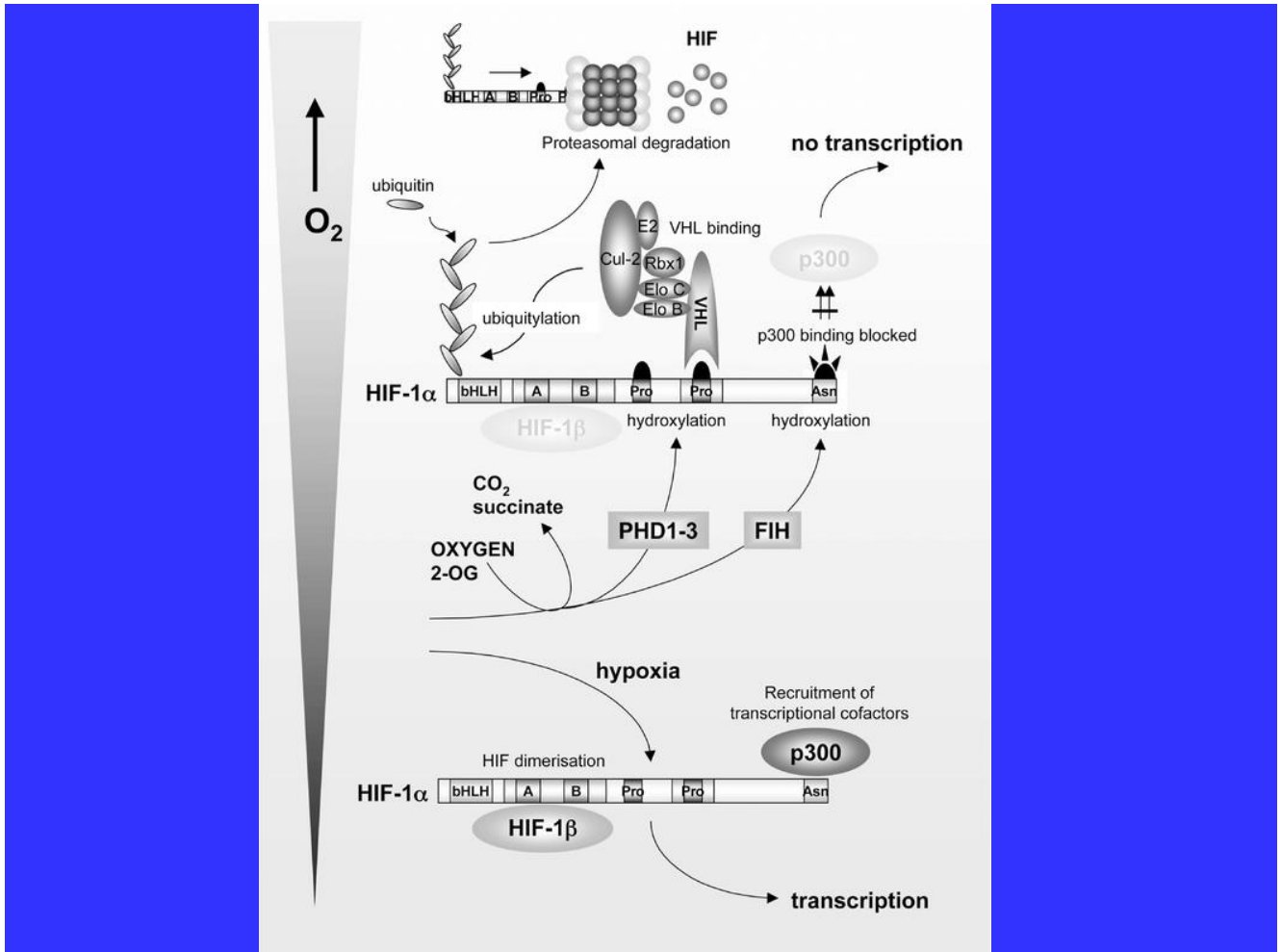


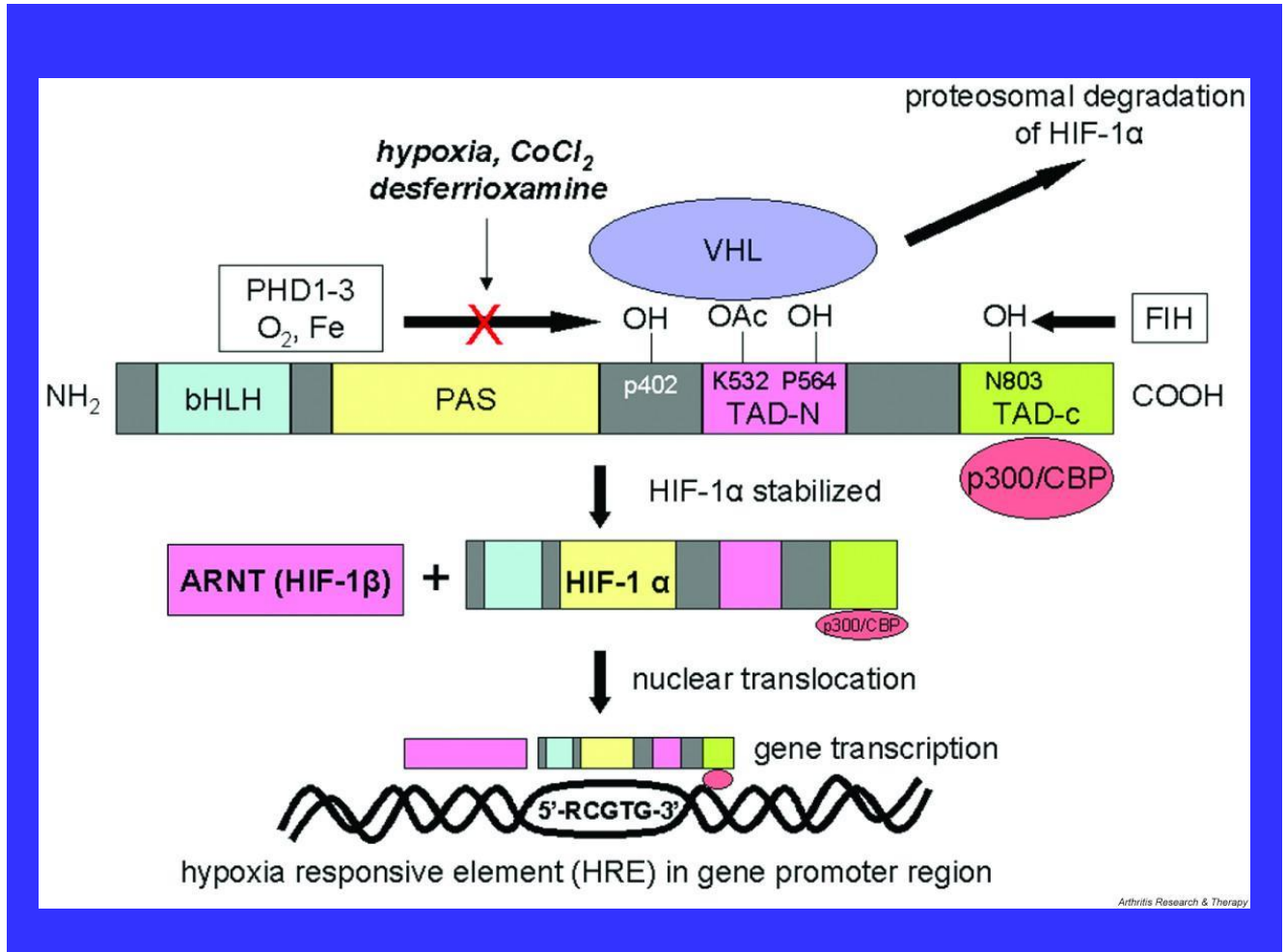
Figure 22–28. Molecular Biology of the Cell, 4th Edition.

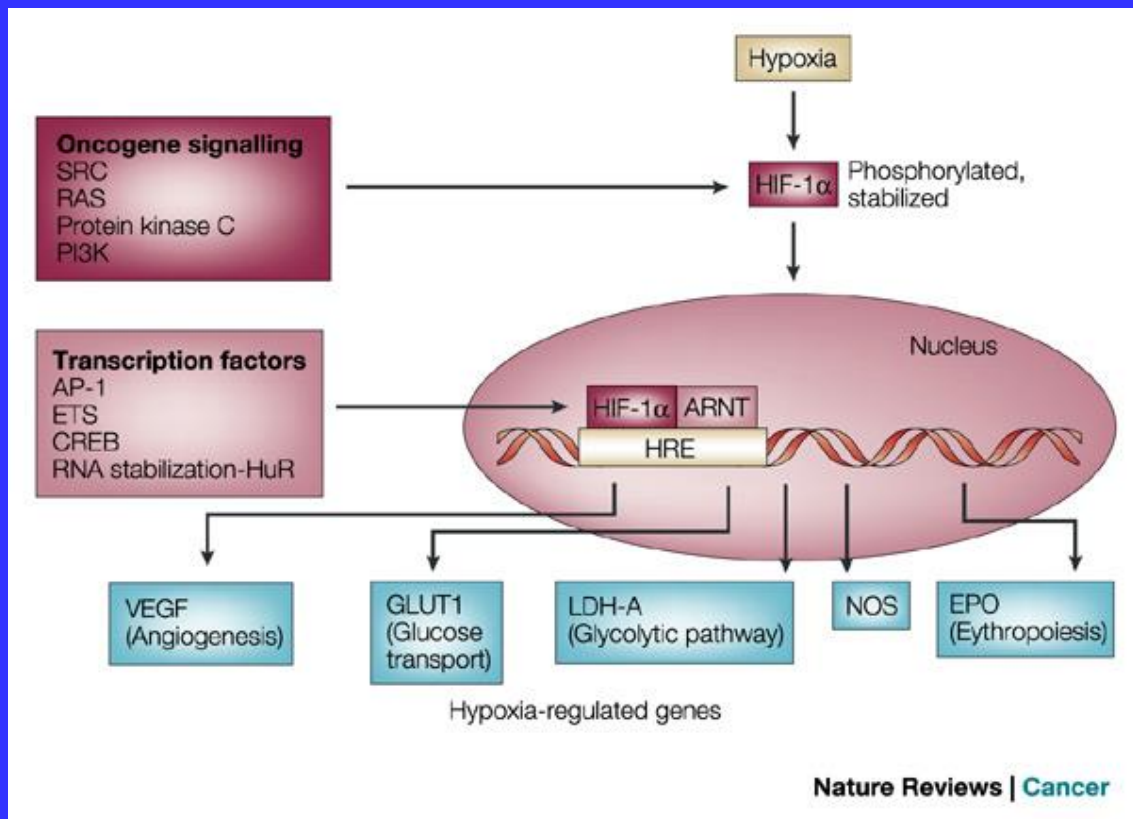
O₂ Sensors

- HIF-1 Prolyl-Hydroxylases 1-3
- Factor Inhibiting HIF-1(FIH1, Asn-hydroxylase)

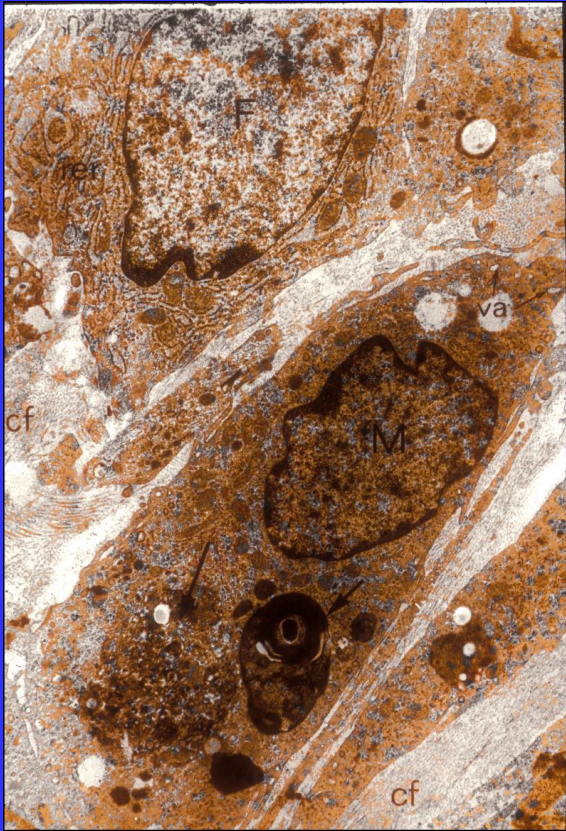


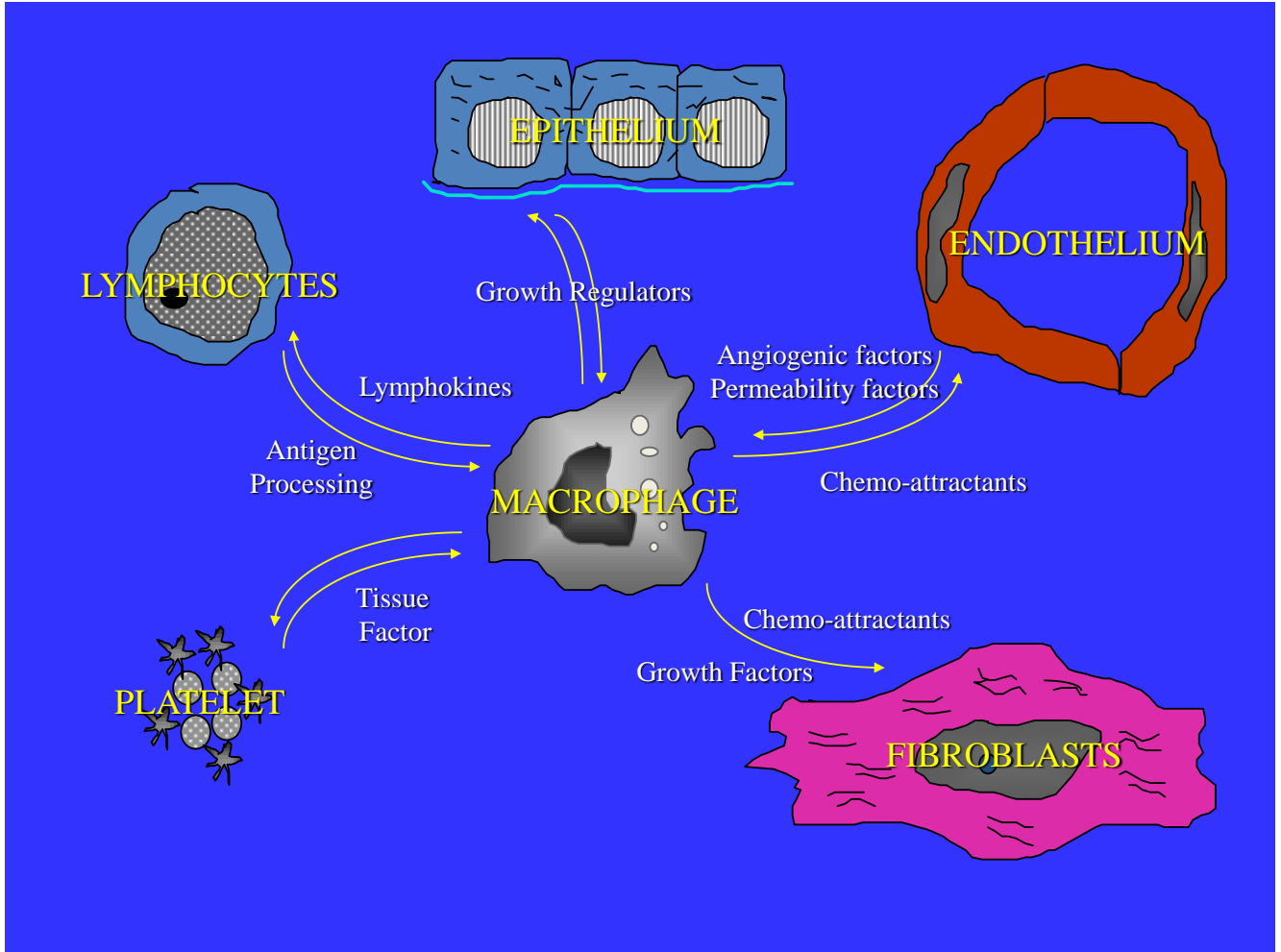




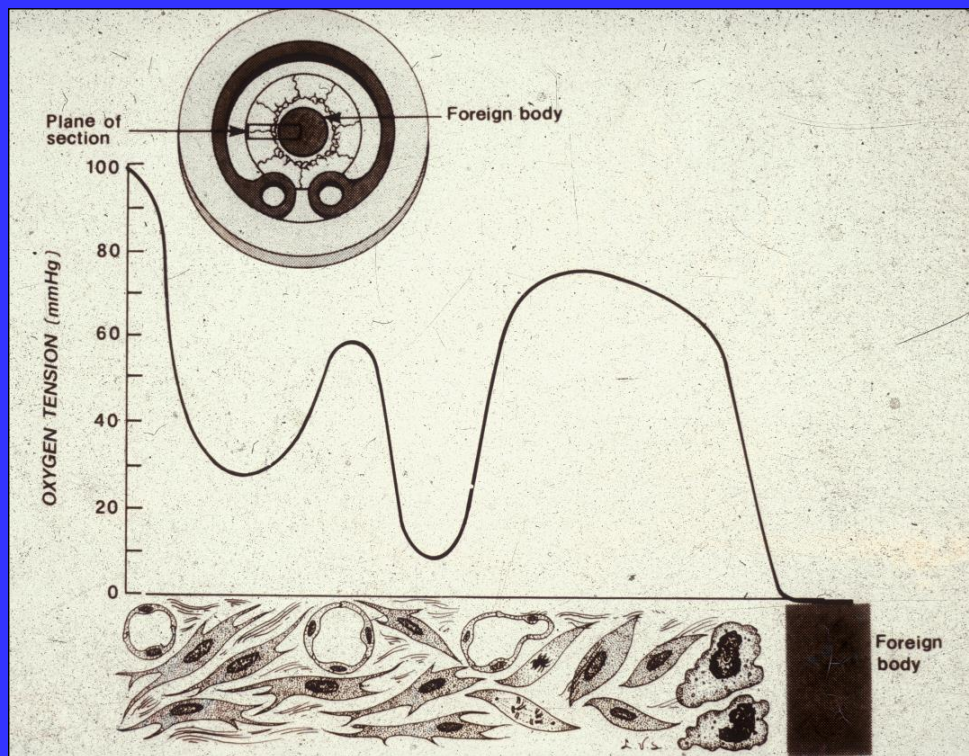


Macrophage in Wound - EM





Role of Hypoxia in Angiogenesis



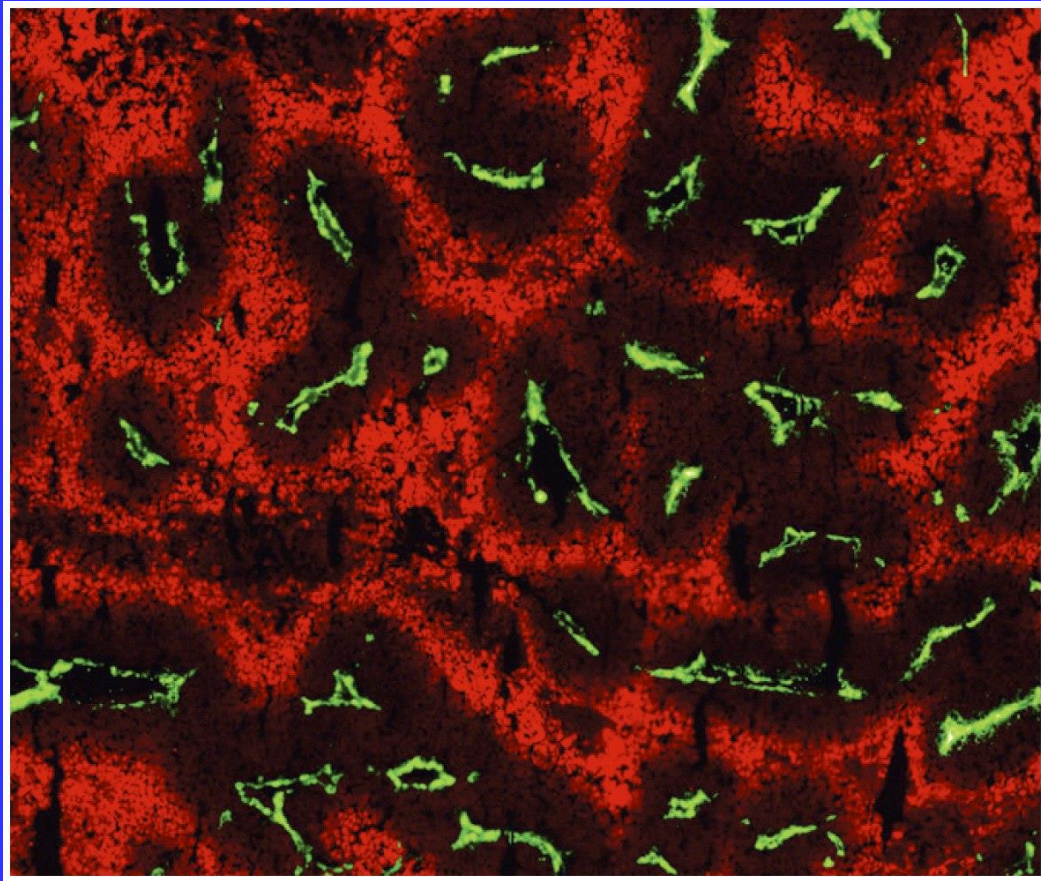
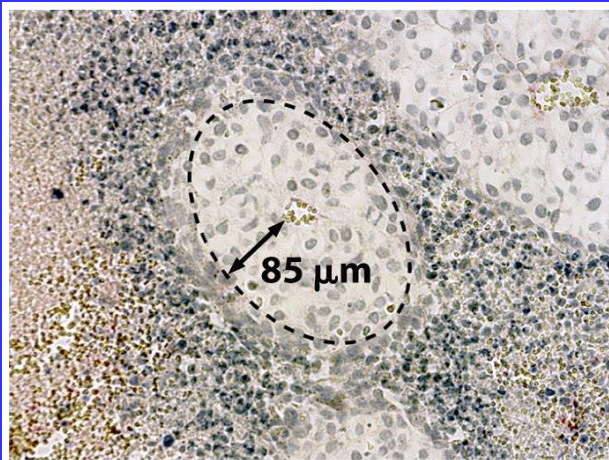
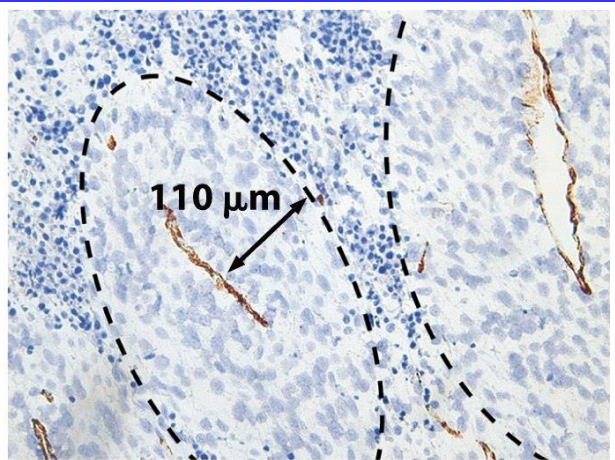


Figure 13.27a *The Biology of Cancer* (© Garland Science 2007)

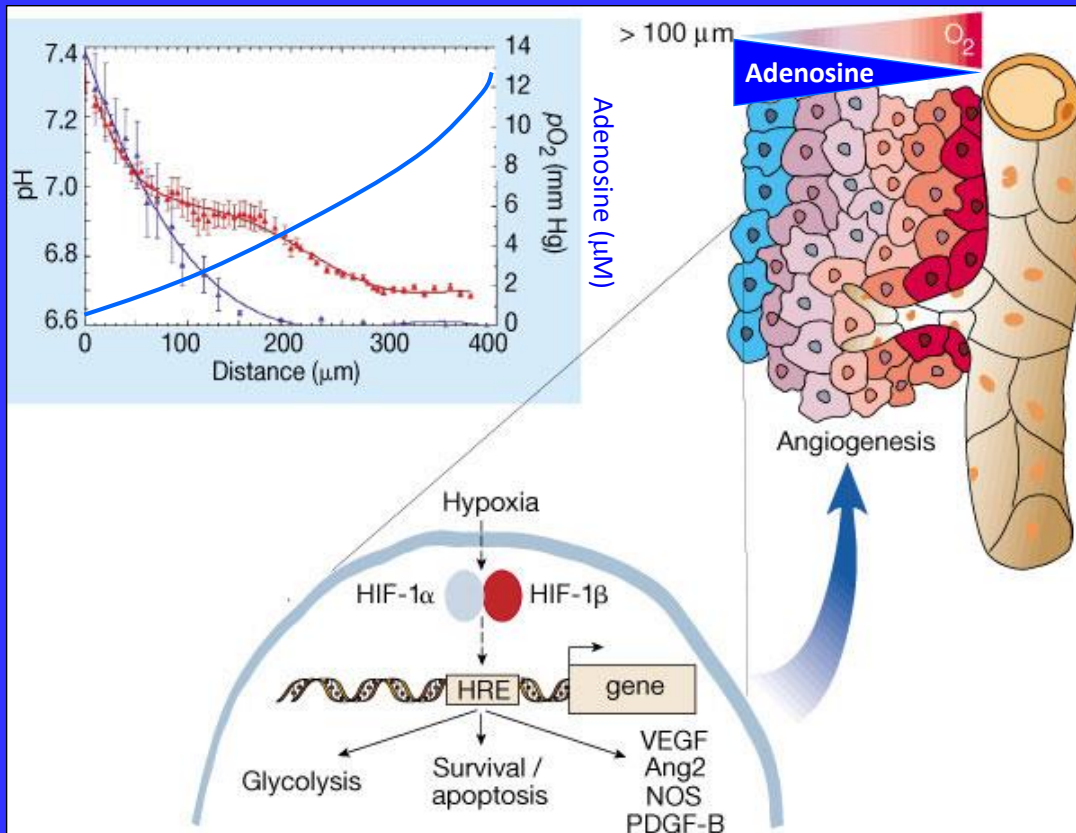


human melanoma

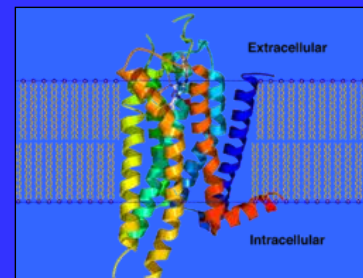
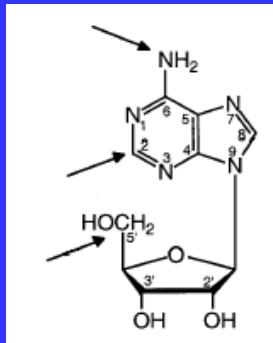


rat prostate cancer

Figure 13.27b *The Biology of Cancer* (© Garland Science 2007)

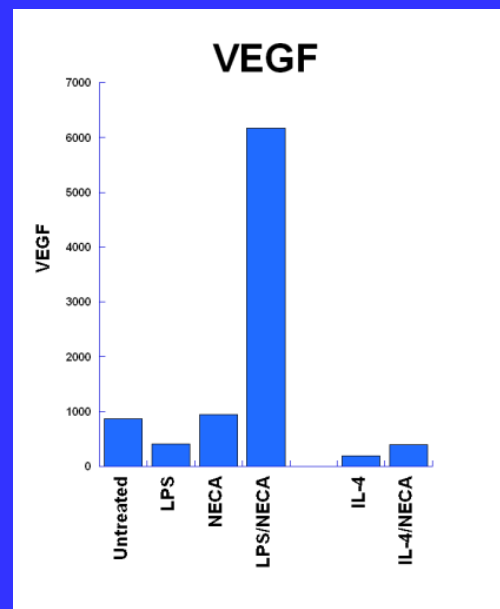
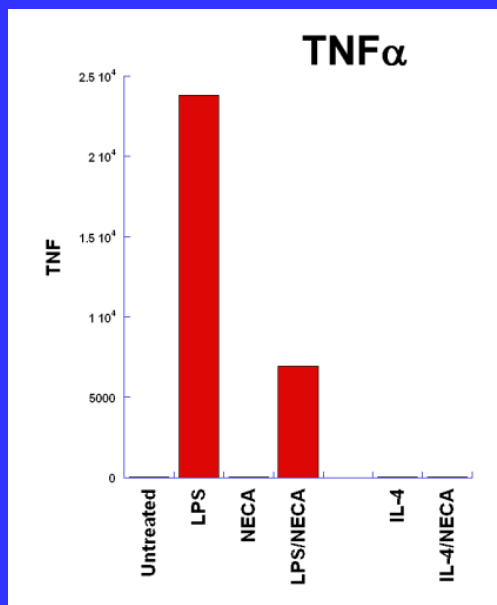


Adenosine and Adenosine Receptors



- Unstable and ubiquitous purine nucleoside produced by breakdown of ATP.
- Released in response to stressful stimuli. Binds receptors to modulate, and protect cells from the harmful consequences of stress
- Adenosine receptors (A₁R, A_{2A}R, A_{2B}R, A₃R) are expressed on many cell types: they bind a wide spectrum of natural and synthetic agonists and antagonists.
- Activation of ARs on immune cells generally exhibits anti-inflammatory effects:
 - Inhibition of phagocytosis.
 - Decreased expression of inflammatory cytokines and chemokines, ROS and NO.
 - Increased expression of anti-inflammatory cytokines.

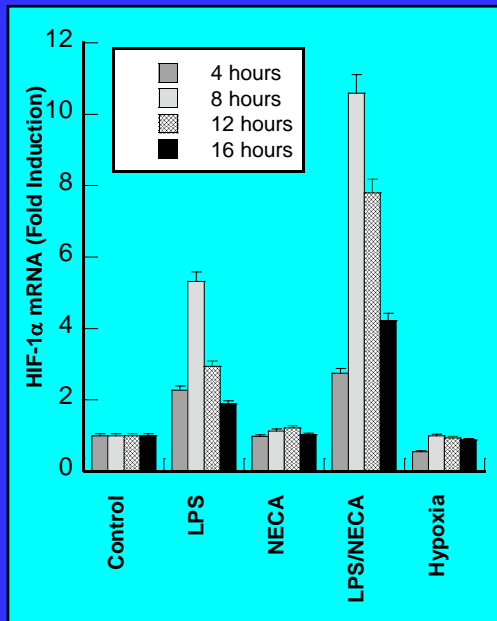
Reciprocal Regulation of TNF α and VEGF in Macrophages In Response to LPS and Adenosine Receptor (AR) Agonists



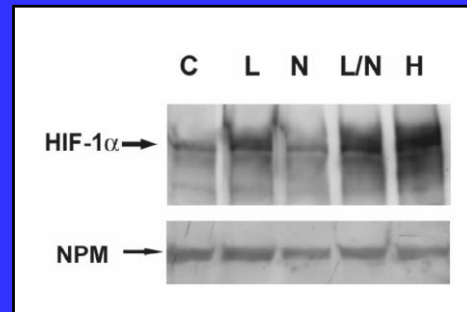
Adenosine synergizes with TLR signaling to switch macrophages from an ***“inflammatory”*** to an ***“angiogenic”*** phenotype

Regulation of HIF-1 α Expression by LPS and NECA

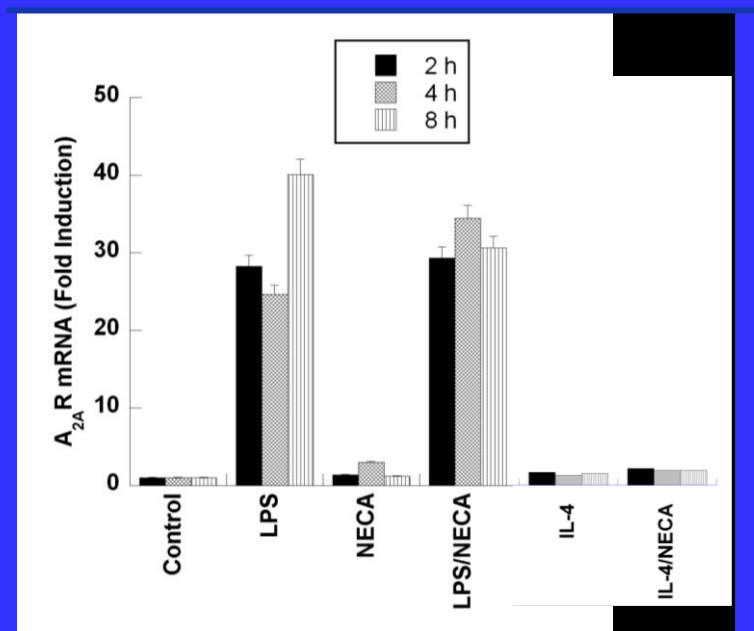
Q-RT-PCR Analysis of HIF-1 α mRNA



Western Blot Analysis of HIF-1 α



Regulation of Adenosine A_{2A} Receptor Expression in Macrophages



- Induction is NF- κ B and STAT1 dependent
- Agonists of TLR2, 7 and 9 also induce A_{2A}R expression

