

RUSH: Seminars in Arthritis and Rheumatism article <# 50130> for proofing by Wallis

=====

Dear Author,

The proof of your article, to be published by Elsevier in Seminars in Arthritis and Rheumatism, is available as a "PDF" file at the following URL:

<http://rapidproof.cadmus.com/RapidProof/retrieval/index.jsp>

Login: your e-mail password

Password: ----

The site contains 1 file. You will need to have Adobe Acrobat Reader software to read these files. This is free software and is available for user download at: <http://www.adobe.com/products/acrobat/readstep.html>

After accessing the PDF file, please:

- 1) Carefully proofread the entire article, including any tables, equations, figure legends and references.
- 2) Ensure that your affiliations and address are correct and complete.
- 3) Check that any Greek letter, especially "mu", has translated correctly;
- 4) Verify all scientific notations, drug dosages, and names and locations of manufacturers;
- 5) Be sure permission has been procured for any reprinted material.
- 6) Answer all author queries completely. They are listed on the last page of the proof;

You may chose to list the corrections (including the replies to any queries) in an e-mail and return to me using the "reply" button. Using this option, please refer to the line numbers on the proof. If, for any reason, this is not possible, mark the corrections and any other comments (including replies to questions) on a printout of the PDF file and fax this to Libby Shuler (717-738-9422) or mail to the address given below.

Do not attempt to edit the PDF file (including adding <post-it> type notes).

Within 48 hours, please return the following to the address given below:

- 1) Corrected PDF set of page proofs
- 2) Print quality hard copy figures for corrections if necessary (we CANNOT accept figures on disk at this stage). If your article contains color illustrations and you would like to receive proofs of these illustrations, please contact us within 48 hours.

If you have any problems or questions, please contact me. **PLEASE ALWAYS INCLUDE YOUR ARTICLE NUMBER** (located in the subject line of this e-mail) **WITH ALL CORRESPONDENCE.**

Sincerely,

Libby Shuler
Issue Manager, YSARH
Cadmus Professional Communications
300 West Chestnut Street
Ephrata, PA 17522
Phone: 717-721-2693
Fax: 717-738-9422
shulerl@cadmus.com



Tumor Necrosis Factor and Granuloma Biology: Explaining the Differential Infection Risk of Etanercept and Infliximab

Robert S. Wallis, MD* and Stefan Ehlers, MD†

Several studies show that the risk of granulomatous infections following therapy with the anti-tumor necrosis factor (TNF) antibody infliximab is higher than after treatment with the soluble TNFRp75 immunoglobulin fusion construct etanercept. Therefore, despite sharing a common target, it is possible that the actual mode of action of the two biologicals differs in vivo. TNF is known to participate in the induction and maintenance of protective granulomas at multiple steps, and evidence supporting a differential inhibition of TNF bioactivity and signaling by the two drugs is discussed.

Semin Arthritis Rheum xx:xxx © 2005 Elsevier Inc. All rights reserved.

Tumor necrosis factor (TNF) is a multipotent cytokine that occurs in a monomeric and trimeric soluble and transmembrane form. The soluble form binds to both TNF receptors (p55 and p75), whereas the transmembrane form predominantly signals via TNFRp75 (1). TNF plays an important role in the pathogenesis of inflammatory diseases such as rheumatoid arthritis (RA) and Crohn's disease (CD). Since 1998, two TNF antagonists have increasingly been used to treat these conditions. Infliximab is a chimeric anti-TNF monoclonal antibody with murine variable regions and human IgG1 constant regions that is administered by periodic infusion. It binds diverse TNF moieties, including monomeric and trimeric soluble TNF and transmembrane TNF (2). Infliximab is effective for the treatment of RA and steroid-refractory CD, in which it induces long-term remissions not readily explained by the kinetics of its inhibition of soluble TNF (3). Case reports indicate activity in sarcoidosis and Wegener's granulomatosis (4,5). In contrast, etanercept is a dimeric fusion protein consisting of the extracellular portion

of the p75 TNF receptor linked to the Fc domains of human IgG1 that is administered twice or once weekly by injection. Etanercept binds only trimeric TNF and interacts with transmembrane TNF with reduced avidity compared with infliximab (2). Etanercept and infliximab are equally effective in RA; however, etanercept is ineffective in CD and sarcoidosis (6,7).

The increased clinical use of TNF antagonists has been accompanied by increased reporting of granulomatous infectious diseases, including tuberculosis (TB), histoplasmosis, and other less common conditions (8). Granulomas represent a host defense strategy to contain intracellular pathogens whose growth cannot be limited by other cellular immune mechanisms (9). Following ingestion of these indigestible pathogens, macrophages release cytokines, chemokines, and other factors that trigger the influx of successive waves of cells to the site of infection (Fig. 1). Initially the response is non-specific, consisting largely of macrophages, neutrophils, and natural killer cells (10). It later progresses to involve lymphocytes with limited antigenic diversity (eg, $\gamma\delta$ T cells) and ultimately results in the recruitment and expansion of highly antigen-specific populations of CD4 and CD8 T-cells. These cells secrete the macrophage-activating cytokines TNF and interferon gamma (IFN γ), express cytotoxicity against infected macrophages, and release antibiotic peptides via granular exocytosis mechanisms. They may also induce apoptosis or otherwise inhibit intracellular microbial growth via other poorly understood mechanisms involving direct cell contact (11,12).

Nonetheless, these mechanisms apparently are not often successful in eradicating *Mycobacterium tuberculosis* infection.

*Associate Professor of Medicine, UMDNJ–New Jersey Medical School, Newark, NJ, USA; Medical Director, PPD Development, Columbia, MD.

†Professor of Medical Microbiology and Immunology of Infectious Diseases, Molecular Infection Biology, Research Center Borstel, Leibniz Center for Medicine and Biosciences, D-23845 Borstel, Germany.

Dr Ehlers has presented data from his own publicly funded research on the role of TNF and lymphotoxin in experimental tuberculosis at several international symposia sponsored by Amgen. Dr. Wallis has served as a consultant for Amgen.

Address reprint requests to Robert S. Wallis, MD, UMDNJ–New Jersey Medical School, Department of Medicine, 185 S. Orange Ave., MSB I-503, Newark NJ 07103 USA. E-mail: r.wallis@umdnj.edu

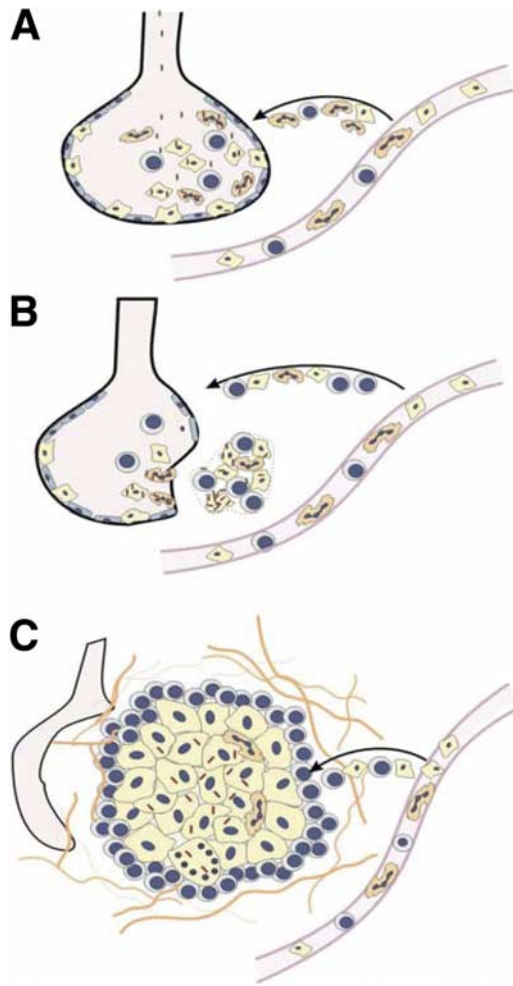


Figure 1 Schematic representation of granuloma induction and maintenance in the lung. (A) Infected macrophages release cytokines and chemokines that induce the recruitment of a mixed (granulocytic and monocytic) cellular infiltrate from the bloodstream into the alveolar and interstitial spaces. (B) The interstitial infiltration becomes predominantly mononuclear in nature and organizes itself into a granuloma with centrally located macrophages and a lymphocytic rim. (C) The fully organized granuloma displaces parenchymal tissue and develops perigranulomatous fibrosis, thereby causing tissue damage. Continuous recruitment of inflammatory cells from the bloodstream is necessary to maintain granuloma structure and contain viable microorganisms persisting in macrophages.

As a result, human mycobacterial immunity is primarily bacteriostatic rather than bacteriocidal, as it results not in sterilization of infected tissues, but in containment of still viable *M. tuberculosis* within granulomas. Maintenance of granulomas is an active process in which cells (both macrophages and T-cells) are continually recruited from the blood (9). Those cells that are not terminally differentiated (such as T-cells) may divide several times in situ before either undergoing apoptosis or returning to the circulation as central memory T-cells. TNF participates in this process at several levels (9,10,13,14). Produced by both macrophages and T-cells, TNF induces antimycobacterial activity in macrophages, promotes the migration of various types of cells to the site of infection, and, under certain circumstances, promotes

apoptosis in T-cells. The relative activities of the soluble and cell-associated forms of TNF in these contexts are not fully understood, with the exception that soluble TNF likely is required to establish the cytokine gradient required to promote cell migration.

Given the multiple sites of TNF action in the host's defense against infections, TNF blockade might be predicted to increase the risk of infections, particularly those normally contained by granulomas. Studies in animal models have indeed provided ample support for this: Mice deficient for TNF or one of its receptors (TNFRp55), or in which TNF function was blocked by neutralizing antibodies, were unable to restrain the growth of microorganisms such as *M. tuberculosis*, *Listeria monocytogenes*, or *Histoplasma capsulatum* (12,15). In the case of mycobacterial infections, these mice showed significantly delayed formation of granulomas, and even the granulomas that formed subsequently disintegrated (16,17). When TNF was experimentally neutralized by antibody after well-organized lesions had been established in wild-type mice, the granuloma structure broke down and dissemination of mycobacteria ensued (18). Similarly, blockade of TNF during chronic latent tuberculosis led to reactivation of mycobacterial multiplication and accelerated death of treated mice (19,20). Because both antibacterially active mechanisms and the demarcation of infectious foci are seriously impaired in the absence of TNF signaling, lesions present as disorganized, diffuse, bacteria-laden, necrotizing infiltrations of mixed cellularity in TNF and TNFRp55-deficient mice.

Unexpectedly, early clinical reports of granulomatous infections appeared to indicate a substantially greater risk posed by infliximab than by etanercept (21-23). A recent report by Wallis and coworkers represents the largest, most systematic study of granulomatous infections associated with infliximab and etanercept contained in the US Food and Drug Administration (FDA) Adverse Event Reporting System (AERS), using reports from 1998 through the third quarter of 2002 (24,25). The number of patients treated with infliximab or etanercept during this time was based on data provided by the manufacturers to the FDA. Of the 35,275 distinct reports extracted from the AERS database arising in the US, 255 described granulomatous infections associated with infliximab and 68 granulomatous infections associated with etanercept. The numbers of treated patients were 197,000 and 113,000, respectively. The overall risk of granulomatous infection was 129 per 100,000 infliximab-treated patients compared with 60 per 100,000 for etanercept ($P < 0.001$). Among the infections that occurred significantly more frequently with infliximab were tuberculosis, histoplasmosis, listeriosis, and coccidioidomycosis; trends toward increased risk were apparent for candidiasis, non-TB mycobacterial disease, and nocardiosis. Infliximab treatment also increased the proportion of extrapulmonary TB cases from 8 to 26% ($P = 0.02$).

Two-thirds of the reports contained data indicating the time from starting TNF antagonist to onset of disease. Time to onset was significantly shorter after starting infliximab for TB (96 versus 350 days, $P < 0.001$) and histoplasmosis (66 versus 518 days, $P = 0.022$); this is consistent with the rec-

ognized importance of reactivation of latent infection in the pathogenesis of these diseases. The risk of TB associated with infliximab was maximal in the first 90 days after starting treatment. If the remaining one-third of TB cases followed a similar distribution, the annualized risk of TB during the first 90 days was 95 cases per 100,000 person-years, compared with 11 per 100,000 person-years for etanercept. For comparison, the reported TB incidence in the US as a whole during this interval was 5 per 100,000 person-years (26).

Thus, despite sharing a common therapeutic target, infliximab is uniquely associated with a high risk of reactivation of latent *M. tuberculosis* infection. This is consistent with the apparent differential therapeutic activities of these two drugs for granulomatous inflammatory conditions and appears to indicate that infliximab, but not etanercept, disrupts established granulomas. Three hypotheses may be proposed to explain these differential effects.

Differential Inhibition of TNF Signaling Events

This hypothesis is based on the differential binding avidities of infliximab and etanercept for soluble versus transmembrane TNF and predicts that inhibition with infliximab would shut down both TNFRp55- and TNFRp75-mediated events, whereas etanercept would leave TNFRp75-mediated signaling at least partially intact (2-27). Since infliximab and etanercept are drugs optimized to bind to human and not mouse TNF, it is difficult to directly address this issue with these reagents in experimental murine models. However, mice deficient in TNF and made transgenic for only the transmembrane form of TNF retained substantial resistance against challenge infections with *M. bovis* BCG or *M. tuberculosis*, arguing that transmembrane TNF signaling via TNFRp75 may provide sufficient antimycobacterial protection under certain experimental conditions (26). In other models of inflammation, host defense mechanisms are severely impaired only in TNFRp55-deficient, but not TNFRp75-deficient, mice (29). In addition, the TNFRp75 seems to be particularly involved in suppressing TNF-mediated inflammatory responses, providing an immunoregulatory feedback loop (29). In corroboration of these findings, studies in murine models of autoimmune diseases have convincingly shown that TNFRp55 signaling is associated mostly with detrimental proinflammatory events, whereas TNFRp75 signaling supports immunomodulatory, disease-ameliorating functions (30).

Differential Power of Neutralizing TNF Bioavailability

Because of its high association and very low dissociation rate, infliximab binds TNF quickly and irreversibly. In contrast, etanercept has both "high-on" and "high-off" binding kinetics, shedding about 50% of soluble TNF and 90% transmembrane TNF within 10 minutes after binding (2). Assuming tissue levels were sufficiently high, infliximab would neutralize all of TNF bioactivity, whereas freely diffusing etanercept would redistribute bioavailable TNF from sites of production to sites of lower concentration.

In the context of granulomatous infections, complete blockade of TNF activity would result in the complete abrogation of inflammatory cell recruitment into the granuloma and a major impairment in macrophage activation. The minimal amount of TNF required to provide sufficient protective functions within the various tissues is of course unknown. However, it is possible that a lower degree of TNF blockade (eg, up to 90%) is still compatible with maintaining both the granuloma structure and the residual antimicrobial macrophage functions in certain circumstances. For example, the rate and success of experimental reactivation of tuberculosis in mice substantially differs depending on the time postinfection, the duration of antimycobacterial treatment, the anti-inflammatory potency of the reactivating regimen, etc., all of which are factors that determine the relative amount and availability of TNF present in the lesion (19,20). This hypothesis therefore argues that a window of opportunity exists in which TNF levels are high enough to sustain the integrity of granulomas but low enough to reduce the activity of some, but not all, chronic inflammatory disease conditions. Assuming that etanercept and infliximab also differ in vivo in terms of their overall efficacy of neutralizing TNF, the differential therapeutic efficacy of these drugs in CD and sarcoidosis might be explained.

Differential Induction of Target Cell Death

Infliximab has been shown to induce apoptosis both in vitro and in vivo, whereas no such reports exist with etanercept. Specifically, infliximab caused apoptosis in monocytes from CD patients, through a caspase-3-dependent pathway, and, in ex vivo studies, increased apoptosis in CD3+ lymphocytes in the lamina propria of colonic biopsies (31,32). Analyses of tissue biopsies in *M. tuberculosis* infected mice following antibody-mediated inhibition of TNF activity also indicated increased apoptotic activity within granulomatous lesions (20), whereas in patients treated with infliximab apoptosis within pulmonary granulomas was apparently decreased (33). If T-cells with specific reactivity against mycobacterial antigens express transmembrane TNF, and if infliximab caused the elimination of these memory cells by inducing apoptosis, reactivation TB would be readily explained by the loss of specifically reactive, IFN γ -producing and macrophage-activating T-cells.

Conversely, a recent phase I study in which etanercept was administered for 1 month as adjunctive treatment to 16 HIV-infected subjects with pulmonary tuberculosis suggested that etanercept may inhibit, rather than promote, T-cell apoptosis (34). The mean baseline CD4 T-cell count was 394 μ L. In the absence of specific antiretroviral therapy, CD4 counts rose in these subjects by 96 cells/ μ L during this treatment, compared with 25 in control subjects ($P = 0.1$). Apoptosis is a recognized mechanism for T-cell depletion in TB and HIV infection; the increased T-cell number observed due to etanercept may be due to its inhibition by etanercept.

It is also possible that infliximab—having bound to membrane-associated TNF on monocytes or lymphocytes—activates complement or causes antibody-dependent cellular cytotoxicity via its Fc tail (35). This would help explain the relatively high occurrence of disseminated TB in infliximab-treated patients, because the lysed granuloma macrophages would spill *M. tuberculosis* organism into the bloodstream. However, no studies have directly demonstrated target cell lysis by infliximab.

Rheumatology has undoubtedly been revolutionized by the use of TNF-targeted biologics, which provide symptom relief even in cases of treatment-refractory chronic inflammatory disorders (36). The unexpected clinical observation that, despite sharing a common therapeutic target, these drugs differ substantially in efficacy and adverse event profiles has encouraged investigations into the true mode of action in vivo, since more than simple neutralization of TNF appears to be involved. This unprecedented interaction between basic and clinical sciences should ultimately lead to the design of a new generation of TNF-targeted drugs in which therapeutic effects can be maximized, and side effects, such as interference with protective granuloma formation, can be minimized.

Acknowledgments

Work in the laboratory of S.E. is supported by grants from the Deutsche Forschungsgemeinschaft (SFB367-C9, SFB415-C7, SFB470-C9, GRK288-A3) and the Focus on Defence against Infection (University of Lübeck, C3). Figure 1 was designed by I. Bouchain. The clinical trial performed by Dr Wallis of etanercept in HIV-associated tuberculosis was partially supported by grants from Immunex and from the National Institutes of Health (NO1-AI45244/AI95383).

References

1. Grell M, Douni E, Wajant H, Lohden M, Clauss M, Maxeiner B, et al. The transmembrane form of tumor necrosis factor is the prime activating ligand of the 80 kDa tumor necrosis factor receptor. *Cell* 1995; 83(5):793-802.
2. Scallon B, Cai A, Solowski N, Rosenberg A, Song XY, Shealy D, et al. Binding and functional comparisons of two types of tumor necrosis factor antagonists. *J Pharmacol Exp Ther* 2002;301(2):418-26.
3. Baert FJ, D'Haens GR, Peeters M, Hiele MI, Schaible TF, Shealy D, et al. Tumor necrosis factor alpha antibody (infliximab) therapy profoundly down-regulates the inflammation in Crohn's ileocolitis. *Gastroenterology* 1999;116(1):22-8.
4. Serio RN. Infliximab treatment of sarcoidosis. *Ann Pharmacother* 2003; 37(4):577-81.
5. Lamprecht P, Voswinkel J, Lilienthal T, Nolle B, Heller M, Gross WL, et al. Effectiveness of TNF-alpha blockade with infliximab in refractory Wegener's granulomatosis. *Rheumatology (Oxford)* 2002;41(11):1303-7.
6. Utz JP, Limper AH, Kalra S, Specks U, Scott JP, Vuk-Pavlovic Z, et al. Etanercept for the treatment of stage II and III progressive pulmonary sarcoidosis. *Chest* 2003;124(1):177-85.
7. Sandborn WJ, Hanauer SB, Katz S, Safdi M, Wolf DG, Baerg RD, et al. Etanercept for active Crohn's disease: a randomized, double-blind,

- placebo-controlled trial. *Gastroenterology* 2001;121(5):1088-94.
8. Weisman MH. What are the risks of biologic therapy in rheumatoid arthritis? An update on safety. *J Rheumatol Suppl* 2002;65:33-8.
9. Saunders BM, Cooper AM. Restraining mycobacteria: role of granulomas in mycobacterial infections. *Immunol Cell Biol* 2000;78(4):334-41.
10. Roach DR, Bean AG, Demangel C, France MP, Briscoe H, Britton WJ. TNF regulates chemokine induction essential for cell recruitment, granuloma formation, and clearance of mycobacterial infection. *J Immunol* 2002;168(9):4620-7.
11. Canaday DH, Wilkinson RJ, Li Q, Harding CV, Silver RF, Boom WH. CD4(+) and CD8(+) T cells kill intracellular Mycobacterium tuberculosis by a perforin and Fas/Fas ligand independent mechanism. *J Immunol* 2001;167(5):2734-42.
12. Stenger S, Hanson DA, Teitelbaum R, Dewan P, Niazi KR, Froelich CJ, et al. An antimicrobial activity of cytolytic T cells mediated by granulysin. *Science* 1998;282(5386):121-5.
13. Flynn JL, Goldstein MM, Chan J, Triebold KJ, Pfeffer K, Lowenstein CJ, et al. Tumor necrosis factor-alpha is required in the protective immune response against Mycobacterium tuberculosis in mice. *Immunity* 1995; 2(6):561-72.
14. Havell EA. Evidence that tumor necrosis factor has an important role in antibacterial resistance. *J Immunol* 1989;143(9):2894-9.
15. Allendoerfer R, Deepe Jr. GS Blockade of endogenous TNF-alpha exacerbates primary and secondary pulmonary histoplasmosis by differential mechanisms. *J Immunol* 1998;160(12):6072-82.
16. Ehlers S, Kutsch S, Ehlers EM, Benini J, Pfeffer K. Lethal granuloma disintegration in mycobacteria-infected TNFRp55-/- mice is dependent on T cells and IL-12. *J Immunol* 2000;165(1):483-92.
17. Ehlers S, Benini J, Kutsch S, Endres R, Rietschel ET, Pfeffer K. Fatal granuloma necrosis without exacerbated mycobacterial growth in tumor necrosis factor receptor p55 gene-deficient mice intravenously infected with Mycobacterium avium. *Infect Immunol* 1999; 67(7):3571-9.
18. Kindler V, Sappino AP, Grau GE, Piguet PF, Vassalli P. The inducing role of tumor necrosis factor in the development of bactericidal granulomas during BCG infection. *Cell* 1989;56(5):731-40.
19. Scanga CA, Mohan VP, Joseph H, Yu K, Chan J, Flynn JL. Reactivation of latent tuberculosis: variations on the Cornell murine model. *Infect Immunol* 1999;67(9):4531-8.
20. Mohan VP, Scanga CA, Yu K, Scott HM, Tanaka KE, Tsang E, et al. Effects of tumor necrosis factor alpha on host immune response in chronic persistent tuberculosis: possible role for limiting pathology. *Infect Immunol* 2001;69(3):1847-55.
21. Fleischmann R. Safety and efficacy of disease-modifying antirheumatic agents in rheumatoid arthritis and juvenile rheumatoid arthritis. *Expert Opin Drug Saf* 2003;2(4):347-65.
22. Fleischmann R, Iqbal I, Nandeshwar P, Quiceno A. Safety and efficacy of disease-modifying anti-rheumatic agents: focus on the benefits and risks of etanercept. *Drug Saf* 2002;25(3):173-97.
23. Gardam MA, Keystone EC, Menzies R, Manners S, Skamene E, Long R, et al. Anti-tumour necrosis factor agents and tuberculosis risk: mechanisms of action and clinical management. *Lancet Infect Dis* 2003;3(3): 148-55.
24. Wallis RS, Broder MS, Wong JY, Hanson JY, Beenhouwer DO. Granulomatous infectious diseases associated with TNF antagonists. *Clin Infect Dis* 2004;38(9):1261-5.
25. Wallis RS, Broder M, Wong J, Beenhouwer ?. Granulomatous infections associated with TNF antagonists: correction. *Clin Infect Dis* 2004; 39(8):1254-1255.
26. Daniel TM, Debanne SM. Estimation of the annual risk of tuberculosis infection for white men in the United States. *J Infect Dis* 1997;175(6): 1535-7.
27. Wajant H, Pfenzenmaier K, Scheurich P. Tumor necrosis factor signaling. *Cell Death Differ* 2003;10(1):45-65.
28. Olleros ML, Guler R, Corazza N, Vesin D, Eugster HP, Marchal G, et al. Transmembrane TNF induces an efficient cell-mediated immunity and resistance to Mycobacterium bovis bacillus Calmette-Guerin infection

- in the absence of secreted TNF and lymphotoxin-alpha. *J Immunol* 2002;168(7):3394-401.
29. Peschon JJ, Torrance DS, Stocking KL, Glaccum MB, Otten C, Willis CR, et al. TNF receptor-deficient mice reveal divergent roles for p55 and p75 in several models of inflammation. *J Immunol* 1998;160(2): 943-52.
 30. Kassiotis G, Kollias G. Uncoupling the proinflammatory from the immunosuppressive properties of tumor necrosis factor (TNF) at the p55 TNF receptor level: implications for pathogenesis and therapy of autoimmune demyelination. *J Exp Med* 2001;193(4):427-34.
 31. Van den Brande JM, Braat H, van den Brink GR, Versteeg HH, Bauer CA, Hoedemaeker I, et al. Infliximab but not etanercept induces apoptosis in lamina propria T-lymphocytes from patients with Crohn's disease. *Gastroenterology* 2003;124(7):1774-85.
 32. Luger A, Schmidt M, Luger N, Pauels HG, Domschke W, Kucharzik T. Infliximab induces apoptosis in monocytes from patients with chronic active Crohn's disease by using a caspase-dependent pathway. *Gastroenterology* 2001;121(5):1145-57.
 33. Keane J, Gershon S, Wise RP, Mirabile-Levens E, Kasznica J, Schwiet-erman WD, et al. Tuberculosis associated with infliximab, a tumor necrosis factor alpha-neutralizing agent. *N Engl J Med* 2001;345(15): 1098-104.
 34. Wallis RS, Kyambadde P, Johnson JL, et al. A study of the safety, immunology, virology, and microbiology of adjunctive etanercept in HIV-1-associate tuberculosis. *AIDS* (in press)
 35. Scallon BJ, Moore MA, Trinh H, Knight DM, Ghayeb J. Chimeric anti-TNF-alpha monoclonal antibody cA2 binds recombinant trans-membrane TNF-alpha and activates immune effector functions. *Cyto-kine* 1995;7(3):251-9.
 36. Stokes DG, Kremer JM. Potential of tumor necrosis factor neutralization strategies in rheumatologic disorders other than rheumatoid arthritis. *Semin Arthritis Rheum* 2003;33(1):1-18.

UNCORRECTED PROOF

63
64
65
66
67
68
69
70
71
72
73
74
75
76
77
78
79
80
81
82
83
84
85
86
87
88
89
90
91
92
93
94
95
96
97
98
99
100
101
102
103
104
105
106
107
108
109
110
111
112
113
114
115
116
117

AUTHOR QUERIES

AUTHOR PLEASE ANSWER ALL QUERIES

1
