

# Arming Pulmonary Macrophages for Pathogen Defense Program Project

## SOP for Tissue harvesting and cell preparation for flow cytometric isolation Version 1.0

### I. Objective of the SOP

The objective of this document is to provide detailed guidelines for preparation of cells for flow cytometry sorting, beginning with dissection of tissues.

### II. Version History

v1.0 Prepared by Amariliz Rivera *et al.* This version is our current stable as of April 2015.

### III. Methods

1. Mice are anesthetized with ~300  $\mu$ L of avertin solution (tribromoethanol 10mg/mL), and exsanguinated.
  - a. Avertin solution
    - i. 20g 2,2,2- tribromoethanol
    - ii. 20 mL 2-methyl-2-butanol
    - iii. ddH<sub>2</sub>O to 1L
2. Lungs are perfused with 10 mL of 1X PBS using a 10 mL luer-lock syringe with a
  - a. 27 ½ gauge needle.
3. Lungs are extracted from mouse, and placed in a collagenase IV solution (Worthington 43N14578).
  - a. Collagenase solution
    - i. 0.071 g of collagenase IV is diluted in 40 mL of 1X PBS.
      1. 5 mL of collagenase solution needed per set of lungs.
    - ii. Solution is filtered through a 0.45  $\mu$ m sterile filter unit PVDF membrane using a 60 mL syringe before use.
4. 1 mL of collagenase solution is placed in a 60mm x 15mm sterile polystyrene plate (1/set of lungs)
5. Lungs are placed in 1 mL of the solution on the plate, and finely minced using a razor blade.
6. 4 mL of collagenase is added for a total of 5 mL of collagenase solution used per set of lungs.
7. Lungs are incubated at 37°C at 5% CO<sub>2</sub> for 45 minutes.
8. Single cell suspension is made by passing the lung several times through a 5 mL luer lock syringe and 20 gauge needle.
9. The single cell suspension is then filtered by placing a funnel lined with 100  $\mu$ m nylon mesh filter paper on top of a new 15 mL conical tube.
10. Centrifuge lung homogenate at 1500 rpm for 5 min.
11. Discard supernatant.
12. Resuspend pellet in 3 mL of Red Blood Cell Lysis Buffer.
  - a. RBC Lysis Buffer pH 7.2-7.4
    - i. NH<sub>4</sub>CL (155mM)
    - ii. NaHCO<sub>3</sub> (10 mM)
13. After proper resuspension, add 5 mL of 1X PBS to neutralize RBC Lysis Buffer
14. Centrifuge lung homogenate at 1500 rpm for 5 min.
15. Discard supernatant.

16. Resuspend pellet in 1 mL of 1X PBS and count.
17. Centrifuge lung homogenate at 1500 rpm for 5 min.
18. Discard supernatant
19. Resuspend pellet at 10 million cells/ mL of FACS Buffer (0.1% sodium azide in 1XPBS) containing Fc Block.
20. Incubate on ice for ~20 min mildly agitating by hand every few minutes to prevent sedimentation of the cells.
21. Fill tube with FACS Buffer and centrifuge sample at 1500 rpm for 5 min.
22. Discard supernatant
23. Resuspend in FACS staining panel (see below)
24. Incubate on ice for ~60 min mildly agitating by hand every few minutes to prevent sedimentation of the cells.
25. Fill tube with FACS Buffer and centrifuge sample at 1500 rpm for 5 min.
26. Resuspend pellet at 10 million cells/ mL in 1% FBS in 1X PBS.
27. Add DAPI to sample.
  - a. DAPI stock is diluted to 5 mg/ mL
  - b. 1  $\mu$ l/ mL is added to sample.
28. Sort (see SOP for sample collection).